

Biotechnology Journal International

20(4): 1-8, 2017; Article no.BJI.33950 ISSN: 2456-7051 (Past name: British Biotechnology Journal, Past ISSN: 2231–2927, NLM ID: 101616695)

Biodegradation of Indeno (1, 2, 3 - c, d) Pyrene and Dibenzo (A, H) Perylene by Aerobic Heterotrophic Bacteria and Cyanobacteria in Brackish Water

Ichor, Tersagh1*, E. M. Aondoakaa¹ and Ande, Sesugh²

¹Department of Biological Sciences, University of Agriculture, Makurdi, Nigeria. ² Department of Chemistry, University of Agriculture, Makurdi, Nigeria.

Authors' contributions

This work was carried out in collaboration between all authors. Author IT designed the project in collaboration with the co-authors. He planned and led the research to execution, analysed data and effected all the corrections as required by the reviewers. Author AS assisted in the design, execution and also contributed in the literature search and statistical analysis. Author EMA was responsible for recording of values during tests and also contributed in literature search. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/BJI/2017/33950 Editor(s): (1) Muhammad Afzal Ghauri, Industrial Biotechnology Division, NIBGE, Incharge University Cell (PIEAS), Faisalabad, Pakistan. (2) Jayanta Kumar Patra, Assistant Professor, School of Biotechnology, Yeungnam University, Gyeongsan, Republic of Korea. Reviewers: (1) Hanan E.-S. Ali, Egyptian Petroleum Research Institute, Egypt. (2) R. Wilfred Sugumar, Madras Christian College, India. Complete Peer review History: http://www.sciencedomain.org/review-history/22945

Original Research Article

Received 5th May 2017 Accepted 12th July 2017 Published 1st February 2018

ABSTRACT

Biodegradation of indeno (1, 2, 3- c, d) pyrene and dibenzo (a, h) perylene; high molecular weight PAHs by isolates of aerobic heterotrophic bacteria, cyanobacteria and its consortium from Boodo Creek characterized with brackish water was monitored for 56 days using GC- MS. The initial concentration of indeno (1, 2, 3- c, d) pyrene in treatment with aerobic heterotrophic bacteria (AHB) was 0.09 mg/l, cyanobacteria (CB) was 0.08 mg/l; AHB+CB was 0.3 mg/l and the control, C was 0.12. Dibenzo (a, h) perylene had AHB 0.10 mg/l; CB 0.07 mg/l; AHB + CB 0.21 mg/l and C 0.23 mg/l. The quantity of the PAH's monitored reduced to 0 on day 56 in all the treatment options though fluctuation in the quantity of the HMW PAH's was observed throughout the period monitored for biodegradation. Biodegradation of indeno (1, 2, 3- c, d) pyrene and dibenzo (a, h) perylene did not vary with time $(p > 0.05)$.

Keywords: Biodegradation; cyanobacteria; Bodo Creek; bacteria.

1. INTRODUCTION

Polycyclic aromatic hydrocarbons (PAHs) are said to be ubiquitous in the environment, mainly due to anthropogenic activities such as combustion or heat related processes that involve hydrocarbons such as coal gasification and other processes like waste incineration [1,2]. PAHs are both persistent in the environment and toxic to humans and other living organisms with several reported to be carcinogenic and mutagenic [3,4]. Environmental Protection Agency has classified indeno (1, 2, 3-c, d) pyrene as a probable human carcinogen among others whereas benzo (g, h, i) pyrene is known for its carcinogenic, mutagenic and teratogenic properties [5].

The solubility of PAHs in water is low hence its bioavailability has made them recalcitrant to microbial attack. In the aquatic environment, PAHs tend to absorb onto the particle phase due to their high hydrophobicity and solid water distribution ratios [6,7,8]. The distribution of PAHs in the aquatic environment is greatly affected by aquatic particulates that act as aggregates of many complicated organic materials [9].

Biodegradation has become the most common remediation method for removing PAH and other pollutants from natural environments [10]. The process is undertaken by a wide variety of bacteria and other microorganisms including cyanobacteria which utilises PAHs as sources of carbon for energy and as part of detoxification process [1].

Several studies have been carried out on the biodegradation of PAHs by indigenous bacteria in natural waters. Degradation of pyrene by bacterial has been reported by a Mycobacterium sp isolated from sediment [11]. Microbial communities in the gulf of Mexico have been reportedly thought to be responsible for intrinsic bioremediation of crude oil released by deep water horizon oil spill causing rapid proliferation of bacterial taxa which decomposes oil hydrocarbons [12,13]. Degradation of a mixture of PAH's; naphthalene, fluorine and benzo(ά) pyrene a highly recalcitrant PAH was carried out and results indicated that all the PAHs studied disappeared from samples collected from the study sites at Orange beach [14]. It was also reportedly degraded by Mycobacterium sp strain

PYR-1 in sediment – water microcosms [15,16]. Previous studies in marine environments have implicated Cycloclasticus as a major PAH degrader among other bacteria [17,18]. Furthermore, most of the literature studies focused on low molecular weight PAHs such as phenanthrene and pyrene, and little is known about the influences of aquatic particles on the biodegradation of high molecular weight PAHs such as chrysene, benzo(a)pyrene and benzo(g,h,i)perylene. It is known that the biodegradation of high molecular weight PAHs, especially those PAHs with more than 4-rings, are always degraded by means of co-metabolism [19]; low molecular PAHs such as phenanthrene could serve as a cometabolic substrate to enhance the biodegradation processes of high molecular weight PAHs [20].

Ideno (1,2,3-cd) pyrene and benzo (g, h, i) perylene are classified as Carcinogenic PAHs among others $[21]$. Ideno $(1, 2, 3 - c, d)$ pyrene is a polycyclic aromatic hydrocarbon which has six fused benzene rings and has been included in the 16 PAHs considered by USEPA as priority pollutant as a result of its acute toxicity, carcinogenicity and teratogenicity [22]. Biodegradation of ideno (1, 2, 3-c,d) pyrene was previously monitored in soils bioaugmented with reeds and immobilized cells of strains and showed high leve of degradation within 40 days in an estuarine reed wetlands stimulator [22]. [23] implicated bacterial cultures of Stenotrophomonas sp, Pandoraea sp and Pseudoxanthomonas mexicana for efficient degradation of ideno (1, 2, 3-c, d) pyrene for 25 days incubation period.

2. MATERIALS AND METHODS

The study aimed at determining the biodegradation potential of aerobic heterotrophic bacteria and cyanobacteria resident in Bodo creek, a brackish water ecosystem to biodegrade benzo (g, h, i) perylene and indeno (1, 2, 3-c,d) pyrene; high molecular weight PAHs.

In the study, molecular characterization of aerobic heterotrophic bacteria and cyanobacteria isolated from petroleum hydrocarbon contaminated Bodo creek was carried out. Investigation of benzo (g, h, i) perylene and ideno (1, 2, 3-c,d) pyrene degradation potential of the isolates was done. To achieve this, different treatment options comprising of aerobic heterotrophic bacteria (AHB), aerobic heterotrophic bacteria and Cyanobacteria (AHB + CB), Cyanobacteria (CB) and the control, (C) were set up in aqueous medium, spiked with known volume of petroleum hydrocarbons and monitored for biodegradation using agilent 7890 model of GC-MS for 56 days as reported in Ichor et al. [1,24]. We hypothesized that the isolated aerobic heterotrophic bacteria and Cyanobacteria from crude contaminated Bodo creek may have adapted to exposure of petroleum hydrocarbons having remained viable and may possess the inherent capability for petroleum hydrocarbon degradation.

3. RESULTS

The results of biodegradation study for indeno (1, 2, $3 - c$, d) pyrene by aerobic heterotrophic bacteria, Cyanobacteria, consortium of aerobic heterotrophic bacteria and Cyanobacteria and the control are as presented in Fig. 1. Its initial

Fig. 1. Biodegradation of Indeno (1, 2, 3- c, d) pyrene

Fig. 2. Biodegradation of Benzo (g, h, i) Perylene

quantity in treatment option AHB was 0.09 mg/l but reduced to 0.02 mg/l and 0.07 mg/l on the first and second week monitored respectively. It reduced to 0 but increased to 0.05 mg/l in week 7 and 0 in week 8 the last day monitored. For CB, the initial quantity was 0.08 mg/l but reduced to 0 on week1 and increased to 0.06 mg/l, reduced to 0 and rose to 0.05 mg/l in week 2,3 and 4 respectively and further reduced to 0 throughout

the remaining weeks monitored. The consortium of AHB+CB had 0.3 mg/l as the initial quantity, 0.1 mg/l and 0.04 mg/l on week 1 and respectively but reduced to 0 and reappeared in week 5 with 0.04 mg/l but reduced and maintained at 0 throughout the remaining period monitored. In the control, C, the initial quantity of ideno (1, 2, 3 – c, d) pyrene was 0.12 mg/l but reduced to 0.03 mg/l and increased to 0.25 mg/l and 0.36 mg/l in week 1, 2 and 3 respectively. It was however reduced to 0 in week 4 and remained at 0 throughout the period monitored.

Fig. 2 shows biodegradation of benzo(g, h, i) perylene by the treatment options employed. The initial quantity for treatment AHB was 0.10 mg/l which was degraded to 0.02 mg/l and 0.07 mg/l on week 1 and 2 but reappeared to 0.05 mg/l on week 7 and reduced to 0 in week 8. For CB, the initial concentration of was 0.07 mg/l which reduced to 0 in week 1 but rose to 0.06 mg/l in week 2. Fluctuation was observed but remained at 0 from week 5 throughout the period monitored. The initial concentration of benzo (g, h, i) perylene for treatment AHB+CB was 0.21 mg/l but was mineralized to 0.09 and 0.04 mg/l in week 1 and 2 respectively but reduced to 0 in week 3 and 4. It rose to 0.04 in week 5 and reduced to 0 throughout the remaining period monitored. The control had initial concentration of benzo(g, h, i) perylene to be 0.23 mg/l which reduced to 0.03 mg/l in week 1 but rose to 0.24 mg/l and 0.35 mg/l in week 2 and remained at 0 throughout the period monitored.

4. DISCUSSION

The study monitored biodegradation of Benzo (g, h, i) Perylene and Indeno (1, 2, 3-c,d) pyrene from crude contaminated waters of Bodo creek; a moderate salt aquatic environment. The aerobic heterotrophic bacteria and cyanobacteria isolates used were reported in Ichor et al. [25]. The isolates and the consortium of the bacteria and cyanobacteria effectively degraded the high molecular weight PAH's used in the study though fluctuations were observed in the course of monitoring the degradation due to novel synthesis as reported in Ichor et al. [24,1]. The result of the present study revealed higher rates of biodegradation of the HMW PAH's tested by the consortium of aerobic heterotrophic bacteria and cyanobacteria compared to other treatments. The overall result provides sufficient evidence on the capability of the resident microbial flora from Bodo creek to efficiently remove HMW PAH's. Mineralization of phenanthrene, fluoranthene, benzo(b) fluoranthene and benzo(k)fluoranthene, anthracene, benzo (a) anthracene and dibenzo(a,h) anthracene by thesame isolates of aerobic heterotrophic bacteria and cyanobacteria interaction in crude oil contaminated brackish water of Bodo creek has been reported previously [24,25,26].

Biodegradation of PAH's in marine environment by bacteria has been reported in previous studies. For example, Cycloclasticus has been frequently detected in marine PAH's degradation [27,28,29,30,18]. At San Diego Bay sediment, bacteria isolates from the genera Cyclosclasticus, Pseudoalteromonas, Marinobacter, Vibrio, Marinomonas and Halomonas grew on chrysene and phenanthrene (28); Porticoccus hydrocarbonoclasticus has been isolated as a PAH degrader [31] and strains of Porphyrobcter and Microbacterium were isolated on benzo (a) pyrene. Degradation of benzo (a) pyrene from marine enrichment was carried out by strains of Stenotrophomonas, Ochrabactrum and Pseudomonas spp [32]. [33] found Sediminicola as a major microbe found in PAH and oil contaminated Liaodong Bay of Bohai Sea in China. Bacillus subtilis BMT4i and Mycbacterium sp PYR degraded benzo (a) pyrene respectively [34,35]; Geobacillus stearothermophilus (AAP7919) was implicated in biodegradation of anthracene [36]; Mycobacterium vanbaalenii PYR-1 degraded phenanthrene, pyrene and dimethylbenz(a) anthracene [37,38]; Pseudomonas putida P15, BS3760 biodegraded Pyrene, phenanthrene, benz(a)anthracene and chrysene [39,40]. 23 (2015) monitored degradation of pyrene and indeno $(1, 2, 3 - cd)$ pyrene by bacterial cells bioaugmented with reeds in an estuarine reed wetland stimulator in small scale natural conditions within 40 days. Its success was linked to its ability to immobilize and enrich PAH degraders, initiate co – metabolism, and ensure PAH bioavailability [41].

Results from previous studies has demonstrated the capability of indigenous cyanobacteria in saline water to degrade PAH's. [42] implicated Agmenellum quadruplicatum in phenanthrene biodegradation; [43] reported 80% removal of phenanthrene by Aulosira fertilissima. [44] reported biodegradation of between 34 to 100% of pyrene by six strains in 7 days from different genera which include; Chlorella, Selenastrum, Chlamydomonas, Scenedesmus and Synechocytis. [45] found rapid degradation in cyanobacterial mats though pure cultures of it could not degrade hydrocarbons. Previous research findings however attributed biodegradation of hydrocarbons to associated heterotrophic bacteria [46,47,48,49] which varies with the result of our present study where pure cultures of the isolated cyanobacteria degraded indeno (1, 2, 3-cd) pyrene and dibenzo (a, h) perylene.

5. CONCLUSION

The study has shown the capability of indigenous microbial community of Bodo creek to biodegrade indeno $(1, 2, 3 - c, d)$ pyrene and dibenzo (a, h) perylene in moderate salt concentrations without bioaugmentation, nutrient amendment and biostimulation. The findings the study has further provided insight in to efficient and effective degradation of high molecular weight PAH's in actual petroleum hydrocarbon contaminated sites and can serve as a model study for stimulating the resident microflora for effective, efficient, quicker bioremediation and restoration of the crude oil polluted aquatic ecosystems of the Niger Delta region of Nigeria.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. Ichor T, Okerentugba PO, Okpokwasili GC. Biodegradation of total petroleum hydrocarbon by aerobic heterotrophic bacteria isolated from crude oil contaminated brackish waters of Bodo creek. Journal of Bioremediation and Biodegradation. 2014;5:5.
- 2. Stroud JL, Paton GI, Semple KT. Microbealiphatic hydrocarbon interactions in soil. J. Applied Microbiol. 2007;102:1239–1253.
- 3. Kweon O, et al. Polycyclic aromatic hydrocarbon metabolic network in Mycobacterium vanbaalenii PYR-1. J. Bacteriol. 2011;193:4326–4337.
- 4. Lafortune I, Juteau P, Déziel E, Lépine F, Beaudet R, Villemur R. Bacterial diversity of a consortium degrading high-molecular-weight polycyclic aromatic hydrocarbons in a two-liquid phase biosystem. Microb. Ecol. 2009;57:455– 468.
- 5. Luch A. The carcinogenic effects of polyclic aromatic hydrocarbons. London Imperial College Press; 2005. ISBN 1-86094-417-5.
- 6. Wang JL, Zang XY, Wang ZT. Environmental microbiology. Higher Education Publisher, Beijing. In Xia et al., (2006). Biodegradation of polcyclic aromatic hydrocarbons in the natural waters of the yellow river: Effects of high

sediment content on biodegradation. Chemosphere. 1988a;65:457-466.

- 7. Wang LS, Zou HX, Han SK. Polycyclic aromatic hydrocarbons analysis methods. Nanjing University Press, Nanjing. Beijing; 1988b. In Xia, et al. Biodegradation of polcyclic aromatic hydrocarbons in the natural waters of the yellow river: Effects of high sediment content on biodegradation. Chemosphere. 2006;65:457-466.
- 8. Johnsen AR, Wick LY, Harms H. Principles of microbial PAH- degradation in soil. Environ. Pollut. 2005;133:71-84.
- 9. Tang HX, Qian Y, Wen XH. Characteristics of organic chemicals in aquatic environment and principles of controlling technique. China Env. Sci. Press, Beijing; 2000. In Xia, et al. Biodegradation of polycyclic aromatic hydrocarbons in the natural waters of the yellow river: Effects of high sediment content on biodegradation. Chemosphere. 2006;65:457-466.
- 10. Vila J, Tauler M, Grifoll M. Bacterial PAH degradation in marine and terrestrial habitats. Curr. Opin. Biotechnol. 2015;33: 95–102.
- 11. Cerniglia CE, Heitcamp MA. Polycyclic hydrocarbon degradation by mycobacterium. Methods in Enzymology. 1990;188:148–153.
- 12. Lu ZM, den Y, van Nostrand JD, Hd He ZI voodeckers J, Zhoa AF, leo Y, Mason JK, D"haesleer P, Hazam TC, Zhou JK. Microbial gene functions enriched in the deepwater horizon deep-sea oil plume. Isme J. 2012;6:451-460.
- 13. Redmond MC, Valentine DL. Natural gas and temperature structured a microbial community response to the Deepwater Horizon oil spill. Proc. Natl. Acad. Sci. USA. 2012;109:20292–20297.
- 14. Kappell AD, Wei Y, Newton RJ, Van Nostrand JD, Zhou J, McLellan SL, et al. The polycyclic aromatic hydrocarbon degradation potential of Gulf of Mexico native coastal microbial communities after the Deepwater Horizon oil spill. Front. Microbiol. 2014;5:205.

DOI:10.3389/fmicb.2014.00205

- 15. Heitcamp MA, Cerniglia CE. PAH degradation by a mycobacterium sp in microcosms containing sediment and water from a pristine ecosystem. Appl. Environ. Microbiol. 1989;55(8):1968–73.
- 16. Kelley L, Cerniglia CE. Degradation of amixture of high molecular weight PAH's

by a mycobacterium strain PYR – 1. Journal of Soil Contamination. 1995;4:77– 91.

- 17. Kasai Y, Kishira H, Haramaya S. Bacteria belonging to the genus Cycloclasticus play a primary role in the degradation of aromatic hydrocarbons released in a marine environment. Appl. Environ. Microbiol. 2002;68:5625–5633.
- 18. Jurelevicius D, Alvarez VM, Marques JM, de Sousa Lima LR, Dias Fde A, Selsin L. Bacterial community response to petroleum hydrocarbon amendments in freshwater, marine and hypersaline water containing microcosms. Appl. Environ. Microbiol. 2013;79:5927–5935.
- 19. Preuss S, Wittneben D, Lorber KE. Microbiological degradation of polycyclic aromatic hydrocarbons: Anthracene, benzo(k)fluoranthene, dibenzothiophene, benzo(h)quinoline and 2 – nitronaphthalene. Toxicol. Environ. Chem. 1997;58: 179–195.
- 20. Gong ZQ, Li PJ, Wang X. Cometabolic degradation of benzo(a)pyrene in the soil by the introduced fungi. Res. Env. Sci. 2001;14(6):36-39.
- 21. Mahmood A, Syed JH, Malik RN, Zheng Q, Chang Z, Li J, Zhang G. Polychlorinated biphenyls (PCBs) in air, soil and cereal crops along the two tributaries, distribution and screening level risk assessment. Science of the Total Environment. 2014;481:596–604.
- 22. Cerniglia CE. Biodegradation of polycyclic aromatic hydrocarbons. Biodegradation. 1992;3:351-368.
- 23. Huang RY, et al. Enhanced biodegradation of pyrene, and ideno $1,2,3 -$ cd pyrene using Bacteria immobilised in cinder Beads in Estuarine wetlands. Mar. Pollut. Bull. 2015;102(1):128–133.
- 24. Ichor Tersagh, Ebah EE, Azua ET. Aerobic of fluoranthene, benzo(b)fluoranthene and benzo(k) fluoranthene by aerobic heterotrophic bacteria cyanobacteria interaction in brackish water of bodo creek. Journal of Petroleum & Environmental Biotechnology. 2016a;7:4.

DOI: 10.4172/2157-7463.1000292.

25. Ichor T, Gberikon GM, Dooshima Nevkaa**.** Biodegradation of phenanthrene by a consortium of aerobic heterotrophic bacteria and cyanobacteria in petroleum polluted brackish water of bodo creek. Microbiology Journal. 2016b;6:1-8. ISSN 2153-0696. DOI: 10.3923/mj.2016.

26. Ichor Tersagh, Aondoakaa EM, Okpa BO. Aerobic degradation of anthracene, benzo(a) anthracene and dibenzo(a,h) anthracene by indigenous strains of aerobic heterotrophic bacteria and cyanbacteria isolates from crude oil contaminated bodo creek. Oil and Gas Res 2017;3:125.

DOI:10.4172/2472-0518.1000125.

- 27. Melcher RJ, Apitz SE, Hemmingsen BB. Impact of irradiation and polycyclic aromatic hydrocarbon spiking on microbial populations in marine sediment for future aging and biodegradability studies. Appl. Environ. Microbiol. 2002;68:2858- 2868.
- 28. Hazen TC, Dubinsky EA, Desantis TZ, Anderson GL, Piceno YM, Singh Njasson, kprobst J, et al. OTU deep sea oil plume enriches indigenous oil degrading bacteria. Science. 2010;330:204-208.
- 29. Chakraborty R, Borglin SE, Dubinsky EA, Andersen GL, Hazen TC. Microbial response to the MC-252 oil and Corexit 9500 in the Gulf of Mexico. Front. Microbiol. 2012;3:357.
- 30. Mason OU, Hazen TC, Borglin S, Chain PSG, Dubinsky EA, Fortney JL, et al. Metagenome, metatranscriptome and single cell sequencing reveal response to Deepwater Horizon. ISMEJ. 2012;6:1715– 1727.
- 31. Guittierez T, Nicholas PD, Whiteman WB. Atkten MB. Porticoccus hydrocarbonoclasticus sp nov., an aromatic hydrocarbon degrading bacterium indentified in laboratory cultures of marine phytoplankton. Appl. Environ. Microbiol. 2012;78:628-637.
- 32. Luo YR, Lai QL, Huang H, et al. Analysis of community structure of microbial consotuim capable of degrading benzo {a}pyrene by the DGGE. Mar. Pollut. Bull. 2009;58:1159-1163.
- 33. Zheng B, Wang L, Liu L. Bacterial community structure and its regulating factors in the intertidal sediment along the Liaodong Bay of Bohai Sea, China. Micobiological Res; 2014. DOI: 10.1016/j.micres.2013.09.019.

Ichor et al.; BJI, 20(4): 1-8, 2017; Article no.BJI.33950

- 34. Lily MK, Bahuguna A, Dangwal K, Garg V. Degradation of Benzo[a]pyren by novel train Bacillus subtilis BMT4i (MTCC9447). Brazilan Journal of Microbiology. 2009;40(4).
- 35. Cheung PY, Kinkle BK. Mycobacterium diversity and pyrene mineralization in petroleum-contaminated soils. Applied Environmental Microbiology 2001;67: 2222-2229.
- 36. Kumar V, Kothiyal NC. Distribution behaviour of polycyclic aromatic hydrocarbons in roadside soil at traffic intercepts within developing cities. International Journal of Environmental Science and Technology. 2011;8(1):63-72.
- 37. Kim TJ, Lee EY, Kim YJ, Cho KS, Ryu HW. Degradation of polyaromatic hydrocarbons by Burkholderia cepacia 2A-12. World Journal of Microbiology and Biotechnology. 2003;19:411-417.
- 38. Lopez Z, Vila J, Minguillon C, Grifoll M. Metabolism of fluoranthene by Mycobacterium sp. strain AP1. Appl. Microbiol. Biotechnol. 2006;70:747–756.
- 39. Kazunga C, Aitken MD. Products from the incomplete metabolism of pyrene by polycyclic aromatic hydrocarbon degrading bacteria. Applied Environmental Microbiology. 2000;66:1917-192.
- 40. Balashova NV, Kosheleva IA, Golovchenko NP, Boronin AM. Phenanthrene metabolismby Pseudomonas and Burkholderia strains. Process Biochemistry. 1999;35:291- 296.
- 41. Yap CL, Gan S, Ng HK. Application of vegetable oils in the treatment of polycyclic aromatic hydrocarbons – contaminated soils. J. Hazard. Mater. 2010;177:28–41.
- 42. Narro ML, Cerniglia CE, van Baalen C, Gibson DT. Metabolism of phenanthrene by the marine cyanobacterium Agmenellum quadruplicatum PR-6. Applied Environ. Microbiol. 1992;58:1351-1359.
- 43. Ji NK, Barot M, Khan SR,. Some intermediate bio-transformants during biodegradation of high molecular weight phenanthrene and fluoranthene by cyanobacterial species-Aulosira fertilissima ghose. Int. J. Applied Sci. Biotechnol. 2013;1:97-105.
- 44. Lei AP, Wong YS, Tam NFY. Removal of pyrene by divergent microagal species. Water Science Technology. 2002;46(12): 195-201.
- 45. Cohen Y. Bioremediation of oil by marine microbial mats. International Microbiology. 5: 189-193 cyanobacterial consortium. Microb. Ecol. 2002;50:500-588.
- 46. Chaillan F, Gugger M, Sailot A. The role of cynobacteria in the biodegradation of crude oil by the tropical cynobacterial mat. Chemosphere. 2006;62:1574–1582.
- 47. Sanchez O, Elia D, Esteve I, Mas J. Molecular characterization of an oildegrading Cyanobacterial consortium. Microbial Ecology. 2005;50:500-588.
- 48. Abed RMM, Koster J. The direct role of aerobic heterotrophic bacteria associated with cyanobacteria in the degradation of oil compounds. International Biodeterioration & Biodegradation. 2005;55:29-37.
- 49. Abed RMM, Safi NMD, Koster J, De Beer D, El-Nahhal Y, et al. Microbial diversity of a heavily polluted microbial mat and its community changes following degradation of petroleum compounds. Applied and Environmental Microbiology. 2002;68: 1674-1683.

___ © 2017 Ichor et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

> Peer-review history: The peer review history for this paper can be accessed here: http://www.sciencedomain.org/review-history/22945