



Repurposing Bazedoxifene for Cancer Therapeutics: It is Much more than a SERM

Tomas Koltai^{1*}

¹Maipu 712, Buenos Aires, Argentina.

Author's contribution

The sole author designed, analysed, interpreted and prepared the manuscript.

Article Information

Editor(s):

(1) Dr. Bing Yan, Hainan Branch of PLA General Hospital, China.

Reviewers:

(1) Ahmed Mohammed Morsy, Assiut University, Egypt.

(2) Mohd Adzim Khalili Bin Rohin, Universiti Sultan Zainal Abidin (UniSZA), Malaysia.

Complete Peer review History: <http://www.sdiarticle4.com/review-history/62890>

Review Article

Received 18 September 2020

Accepted 23 November 2020

Published 11 December 2020

ABSTRACT

Bazedoxifene (BDF) is a selective estrogen receptor modulator (SERM) that has been approved by the FDA for the treatment of post-menopausal osteoporosis in association with conjugated estrogens. BDF shares many of the pharmacological effects of tamoxifen with the advantage of not being an ER agonist in uterus and decreasing the risk for endometrial carcinoma induced by tamoxifen.

Interestingly, BDF has shown anti-tumoral actions in tissues and tumors that are hormone-independent. This means that BDF is not only a SERM. The better known of these mechanisms are

- 1) Inhibition of the IL6-IL6R-GP130-STAT 3 axis (IL6: interleukin 6; IL6R: interleukin 6 receptor; GP130: glycoprotein 130; STAT3: Signal Transducers and Activators of Transcription-3).
- 2) Modulation of the Hippo-YAP pathway.
- 3) Inhibition of AOX1 (aldehyde oxidase 1)

The first two are neatly anti-tumoral. The third one is sort of controversial.

This review is focused on the non-hormonal anti-tumor mechanisms of BDF. Its repurposing for the treatment of malignancies, other than breast cancer, is analyzed.

Keywords: *Bazedoxifene; SERM; interleukin 6; interleukin 6 receptor; GP130; STAT3; breast cancer.*

*Corresponding author: E-mail: tkoltai@hotmail.com;

1. INTRODUCTION

At the end of the XIX century the Scottish surgeon George Beatson, based on the observation of the ovarian influence on the breast of animals, removed both ovaries in three patients with breast cancer. To his surprise all the three improved. He had no idea about hormones or estrogens [1], but unknowingly the first step in the relationship between estrogens and breast cancer started to be disclosed.

At the beginning of the 1980s a firm suspicion on the role of estrogen replacement therapy in breast cancer developed in the scientific community [2,3]. All the evidence showed that hormone replacement therapy in postmenopausal women increased the risk of breast cancer and that estrogen depletion could be an important mechanism for its treatment. Research was oriented towards the discovery of an anti-estrogen compound. Tamoxifen, not being the first, was found to be quite effective as an anti-estrogen in breast [4].

On October 2013 the Food and Drug Administration (FDA) approved bazedoxifene (Bazedoxifene CAS#198481-32-2). The FDA approved the new medication Duavee®, comprised of Bazedoxifene (BDF) and conjugated estrogens as a preventive treatment for postmenopausal flushes and osteoporosis.

BDF has been previously approved by the European Medicines Agency in 2009. The addition of BDF to the conjugated estrogens decreases excessive growth of the uterine mucosa, lowering uterine cancer risk due to estrogens. Leaving aside the conjugated estrogens, our analysis will be limited to BDF as a possible anticancer drug.

BDF is an indole derivative (Fig. 1) acting as a selective estrogen receptor modulator (SERM) and selective estrogen receptor degrader (SERD) with mixed agonist and antagonist actions on the estrogen receptor (ER) according to tissue specificity.

It has ER antagonistic effects on breast and uterus, while tamoxifen being an antagonist in breast is an agonist in uterus. BDF binds the estrogen receptors (ER) in responsive tissues, such as breast, endometrium, bone, and liver. BDF bound to ER translocates to the nucleus where it may act as an antagonist in breast and endometrium blocking the effects of estrogen-ER complex. It can also act as an agonist in liver, modifying lipid metabolism decreasing total and LDL cholesterol [5]. Effects on bone are mainly anti-resorption increasing mineral density. BDF's main pharmacological characteristic is its tissue specificity. It is an estrogen antagonist in some tissues and at the same time an agonist in others.

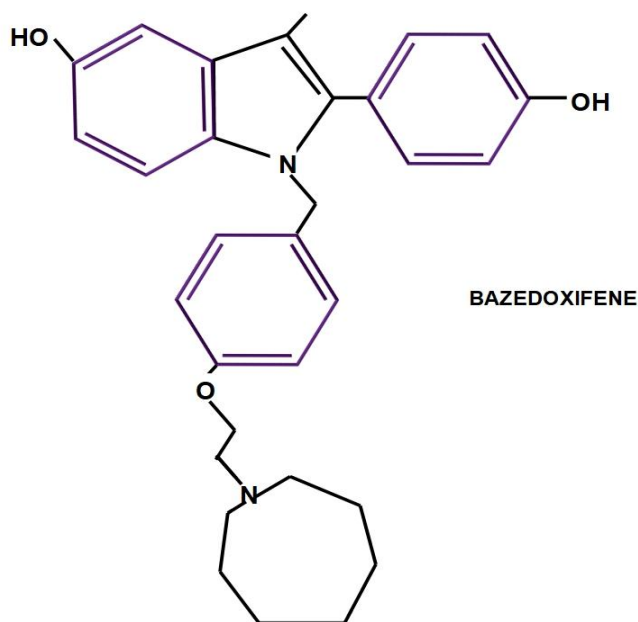


Fig. 1. Chemical structure of the hybrid SERM/SERD bazedoxifene

BDF is used in the following situations:

- 1) Prevention and/or treatment of breast cancer [6].
- 2) Prevention and/or treatment of post-menopausal osteoporosis [7,8,9,10].
- 3) Treatment of menopausal symptoms such as hot flashes (associated with conjugated estrogens) [11,12,13].

The objective of this review is to analyze BDF for the treatment of cancer in general, including but not exclusively breast cancer. The other two indications, namely osteoporosis and hot flashes will not be considered here. (Reviewed by Peng et al. [14], Banzal et al. [15], and Parish et al. [16])

2. NON-HORMONAL ANTICANCER ACTIONS OF BAZEDOXIFENE

The mechanism of action of bazedoxifene in non-hormonal cancer has not been fully cleared, but evidence has been accumulating in this sense. It is a SERM and a SERD, but acts also on hormone-independent breast cancer [17] showing that there must be other actions involved.

In the first place, as a SERM it has effects that are superior to that of tamoxifen rendering it useful in the treatment of tamoxifen-resistant breast cancer [18,19]. Resistance to tamoxifen is acquired through mutations of the estrogen receptor alpha (ER α). BDF has the ability to degrade these mutant forms of ER α [20], therefore it is also a SERD. These mutations are quite common after cancer treatment [21].

BDF potentiated anti-tumoral action of paclitaxel in breast cancer, increasing apoptosis, decreasing cell viability, migration and colony formation [22]. It was shown to be an inhibitor of triple negative breast cancer cells growth. This was accomplished by an independent activity from ER through inhibition of the IL6-GP130 pathway [23,24,25]. The inhibition of this axis has an anticancer role in breast cancer and also in other hormonal-independent tumors.

Three BDF effects related with non-hormonal cancers were clearly identified:

- 4) Inhibition of the IL6-IL6R-GP130-STAT 3 axis (IL6: interleukin 6; IL6R: interleukin 6 receptor; GP130: glycoprotein 130; STAT3: Signal Transducers and Activators of Transcription-3.

- 5) Modulation of the Hippo-YAP pathway.
- 6) Inhibition of AOX1 (aldehyde oxydase 1)

2.1 Inhibition of the IL6-IL6R-GP130-STAT 3 Axis

The mechanisms involved in this pathway are:

- 1) binding of IL-6 to the IL-6 receptor (IL-6R);
- 2) thus inducing activation of the receptor with
- 3) homodimerization and recruitment of glycoprotein 130 (GP130);
- 4) while IL-6 R is not a signaling molecule, GP130 is the actual signaling glycoprotein [26];
- 5) this pro-tumoral signaling plays a role in growth and metastasis of breast cancer cells [27] and other tumors.

STAT3 is one of the main downstream protein targets, through the JAK-STAT axis. Fig. 2.

As shown in Fig. 2 there are many pro-tumoral pathways initiated by the signaling of IL6-IL6R besides the JAK-STAT pathway. Therefore, IL6-IL6R-GP130 can be considered an oncogenic cytokine hub.

Fig. 2 shows membrane-bound IL6R activation and signaling. However, there is a second mechanism involved that seems to be at least, or more important than membrane-bound IL6R activation. This is the activation of the soluble form of IL6R.

Soluble IL6R (sIL6R) has two origins [28]:

- a) alternative splicing of the IL6R primary transcript [29];
- b) limited proteolysis and shedding of the membrane bound IL6R [30].

Uniprot [31] says that "the restricted expression of the IL6R limits classic IL6 signaling to only a few tissues such as the liver and some cells of the immune system. Whereas the binding of IL6 and soluble IL6R to IL6ST stimulates 'trans-signaling' " [32]. It is GP130 that explains the pleiotropic effects of cytokines like IL-6. BOX1. This explains how IL6 is able to produce effects in cells lacking IL-6R. This process has been called transsignaling. The common intermediate signaling molecule in both cases is GP130. Blocking of IL-6 impedes both mechanisms leading to GP130 signaling, namely, classic membrane IL-6R binding and transsignaling.

BOX 1: TWO MECHANISMS OF IL-6 ACTIVITY
 1) Binding of membrane bound IL-6R and signaling through GP130
 2) Binding to soluble IL-6R and signaling through membrane bound GP130

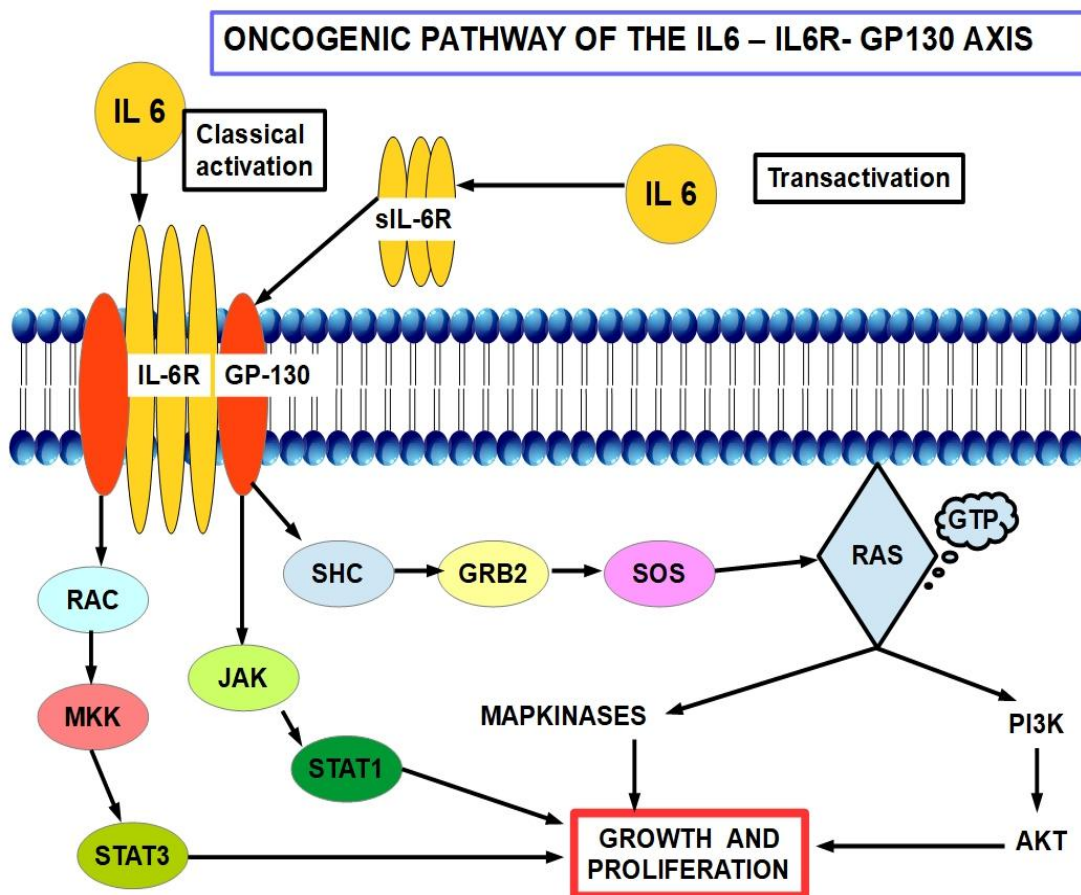


Fig. 2. Oncogenic pathways of the IL6-IL6R-GP130-STAT3 axis at the membrane level. This figure is based on references [33,34,35,36,37,38,39,40,41,42], IL-6 is a cytokine that affects the immune system, inflammation, hematopoiesis, and oncogenesis. It regulates cell growth, gene activation, proliferation, survival, and differentiation. Signaling of IL6 requires the IL6 receptor and GP (Glycoprotein) 130, Binding of IL-6 to its receptor initiates cellular events including activation of JAK (Janus Kinase) and Ras-mediated signaling. Activated JAK phosphorylates and activates STAT transcription factors, particularly STAT3. Activated STAT3 translocates into the nucleus to activate transcription of genes containing STAT3 response elements. STAT3 is essential for GP130-mediated cell survival and G1 to S cell-cycle-transition signals. STAT3 was found to be constitutively activated in many tumors [43]. The Ras-mediated pathway activates MAP kinases downstream. Also PI3K-AKT axis is activated. (PI3K: phosphoinositol 3 kinase; AKT: protein kinase B)

The IL6-IL6R-GP130-STAT3 axis has been described by Chen et al. [44] as a pro-tumoral “vicious” forward cycle driving the tumor, in which IL6 induces STAT3 expression and STAT3 increases IL6. BOX 2

BOX 2
 IL6→IL6R→GP130→STAT3---→IL6....

This pathway has been found to be active in pancreatic cancer and BDF has been proposed as part of the treatment of this cancer, where it seems to down-regulate the IL6/PG130/STAT3 pathway [45,46,47,48]. In pancreatic cancer, the evidence indicates that the anti-cancer mechanism is independent of the hormonal effects of BDF on the ERα. The IL6-GP130-STAT3 signaling axis seems to be an important

tumor driver in many cancers [49,50,51,52,53], including pancreatic cancer. BDF has the ability to disrupt this axis by interfering the IL6R-GP130 relationship, thus blocking GP130 signaling.

This axis is also operative in other tumors where BDF decreased their growth, such as rhabdomyosarcoma [54], head and neck cancer [55], triple negative breast cancer [56], gastrointestinal cancer [57] including colon cancer [58] and gastric cancer [59], medulloblastoma [60], osteosarcoma [61], and hepatocellular carcinoma [62], among others.

Down-regulation of the IL-6R-GP130 signaling has shown synergy with classical oncological treatments such as 5-fluorouracyl in colon cancer [63], paclitaxel in breast cancer, and targeted treatments like lapatinib in breast cancer [64], and palbociclib also in metastatic breast cancer [65].

The persistence of high GP130-STAT signaling in human rhabdomyosarcoma was found to be involved in resistance to cisplatin, doxorubicin and MEK inhibitors [66]. BDF suppressed the resistance.

There is no evidence of BDF actions on human lung cancer. However, in a *Drosophila* lung tumor model BDF showed anti-tumor properties and synergism with afatinib [67].

2.2 Modulation of the Hippo-yap Pathway

Hippo is a protein kinase that controls growth, proliferation and apoptosis through a phosphorylation cascade that inactivates Yes-associated protein (YAP). Mutations of Hippo produce a loss of control of organs growth/size through the unrestricted activity of YAP [68].

There is evidence of its important oncogenic effects in Ewing sarcoma [69], glioblastoma [70], cervix cancer [71], among others. In glioblastoma BDF had the ability to penetrate the blood brain barrier and showed synergistic anti-tumoral effects with paclitaxel. Yap (yes-associated transcription factor coactivator) signaling promotes migration and progression in glioblastoma [72] and in malignant tumors in general [73]. It has not been clearly established the level at which BDF interferes with the Hippo-Yap pathway.

2.3 Inhibition of AOX1

Aldehyde oxidase 1 is an enzyme that intervenes in the detoxification process of aldehydes and the production of ROS (reactive oxygen species) such as hydrogen peroxide (H₂O₂) and superoxide [74]. It is mainly expressed in liver, kidney, adrenal gland, and to a lesser extent in ovary, testis, prostate and pancreas.

Chen et al. found that BDF, but also some other SERMS had an inhibitory effect on AOX1 [75].

AOX1 is involved in the development of colon cancer [76]. However, the role of AOX1 in cancer is controversial. For example, down-regulated expression of AOX1 promoted bladder cancer progression [77].

2.4 Reduction of VEGF, VEGFR2 and COX2

Hou et al. [78] found in a rat endometriosis model that BDF had the ability to reduce proteins related with angiogenesis such as VEGF, VEGFR2 and COX2. There is no proof that this happens also in human tumors. These effects may be hormone-dependent, however, the authors also found concomitantly decreased levels of IL1, IL2, IL6, and TNF α . Thus, the reduction of VEGF, VEGFR2 and COX2 can be a consequence of the IL6 pathway inhibition.

2.5 Aryl Hydrocarbon Receptor Activation

BDF is able to activate the aryl hydrocarbon receptor (AHR) in breast cancer cells whether ER α positive or negative [79]. AHR is a cytosolic transcription factor regulating stem cell maintenance and cell differentiation and is activated by different natural and synthetic ligands. Ligand-activation of AHR allows its translocation into the nucleus where it regulates specific genes, many of them related with metabolism [80]. AHR is involved in the regulation of many pathways related with proliferation, cell motility and differentiation [81]. AHR participates in different stages of carcinogenesis and its progression [82]. The results of AHR activation is a controversial issue, because it produces pro- and anti-tumoral effects. And to further complicate the issue, effects may differ according to the activating ligand. A group of substances, such as MCDF (6-methyl-1,3,8-trichlorodibenzo-*p*-dioxin), are at the same time AHR agonists and exert antiestrogenic activity [83]. BDF probably

belongs to this group where the AHR agonism is associated with antiestrogenic effects.

3. RENAL CELL CARCINOMA

There are no publications about the use of BDF in renal cell carcinoma (RCC). In male rats, the prolonged BDF administration caused renal tubular tumors [84]. However, BDF was not found to be genotoxic [85]. A high serum level of IL6 was predictive of recurrence in RCC [86].

4. DISCUSSION

Bazedoxifene is a SERM and a SERD that is useful for the treatment of breast cancer through its blocking of the ER α . Besides this hormonal action it showed other anti-cancer non hormone-dependent effects.

The IL-6/GP130/STAT3 signaling pathway is essential for the survival and progression of some cancers. Furthermore, a well known oncoprotein like STAT3 is activated by this signaling pathway in such a manner that a forward loop between IL-6-STAT3-IL6 has been found. We may say that these tumors are IL-6 dependent and IL6 driven. In these kind of tumors BDF has shown efficacy as an associated drug to classical chemotherapies or targeted treatments.

BDF is an effective inhibitor of the IL6 signaling pathway. Therefore, its usefulness goes beyond hormone-dependent tumors.

BDF is not a stand alone drug. It needs to be associated with other chemotherapeutic drugs. It also showed ability to reverse multiple drug resistance in heavily treated cases.

However, there are no ongoing experimental protocols with this drug in cancer treatment other than breast. (Clinicaltrials.gov has 48 registered studies as of October 2020. Most of them are for menopausal symptoms and osteoporosis. Six studies are for breast cancer. There are no studies for other type of cancers).

In patients with high serum IL-6 levels a correlation was found between clinico-pathological features, survival and IL-6 level. This has been confirmed in many types of malignant tumors, such as prostate [87,88,89], colorectal [90,91,92], lung [93], breast [94], gastric [95], ovary [96], head and neck [97], among others.

Does this mean that these tumors are IL-6/GP130/STAT3 driven?

Probably not, in many cases. It is not possible to determine if the serum IL-6 increase is causally related with the tumor [98] Furthermore, it can be increased by hypoxia, whether generalized or at the tumor level [99,100,101,102]. However, it is evident that in these increased serum IL-6 tumors the cytokine is playing an important role and it is not an innocent bystander.

IL-6 increase at the tumor level, whether cause or consequence needs to be taken care of because it has notorious pro-tumor activities besides the IL-6/IL-6R/GP130/STAT3 pathway. Some of these pro-tumoral effects are:

- 1) promotion of epithelial-mesenchymal transition [103,104];
- 2) producing immuno evasion through phosphorylation of programmed death ligand-1 (PDL-1) [105];
- 3) promotion of migration, invasion and metastasis [106,107,108];
- 4) contribution to chemoresistance [109,110];
- 5) promotion of stem cell like properties [111,112];
- 6) contribution to cancer progression through the osteopontin-NF-kB pathway [113];
- 7) inducing an inflammatory pro-tumoral environment [114,115];
- 8) stimulating angiogenesis [116] by
 - a) increasing vascular endothelial growth factor (VEGF) [117,118],
 - b) stimulating circulating blood-derived progenitor endothelial vascular cells[119], and
 - c) basic fibroblast growth factor (bFGF) induction [120];
- 9) contribution to cancer cachexia [121,122,123];
- 10) promotion of myeloid-derived suppressor cells [124];
- 11) promotion of the cyclooxygenase 2 (COX2)/prostaglandin E2 (PGE2) / β -catenin signaling [125].

BDF on the other hand is a non-toxic drug that can be added to any treatment protocol without enhancing patients toxicity with the added value of its synergy with classical or targeted treatments. Importantly, BDF administered at usual doses lacks toxicity [126,127,128, 129,130,131].

The lack of clinical trials on this matter should change and BDF should come into the

therapeutic perspective once its benefits are further confirmed with well planned prospective clinical trials.

5. CONCLUSIONS

There are a group of tumors in which the IL-6/GP130/STAT3 is a driver pathway. In these tumors the association of the non-toxic SERM BDF should improve treatment results. The difficulty lies in determining when a tumor is driven by this pathway. At this stage of our knowledge the serum IL-6 level is not a proof in this sense. Probably, those tumors with normal serum IL-6 levels are not driven by the IL-6 pathway. However, the tumors that express high IL-6, GP130 and STAT3 in the specimen should be considered for associating BDF to the treatment protocol. There is evidence that BDF can be beneficial also in tumors that are not driven by the IL-6/GP130/STAT3.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

COMPETING INTERESTS

Author has declared that no competing interests exist.

REFERENCES

1. Mukherjee S. The emperor of all maladies. A biography of cancer. Scribner. New York. 2010;214.
2. Dukes MNG. Sex hormones and related compounds, including hormonal contraceptives. In Side Effects of Drugs Annual. Elsevier. 2011;33:851-880. Available: <https://www.sciencedirect.com/science/article/pii/B9780444537416000404>
3. Steinberg KK, Thacker SB, Smith SJ, Stroup DF, Zack MM, Flanders WD, Berkelman RL. A meta-analysis of the effect of estrogen replacement therapy on the risk of breast cancer. *Jama*. 1991; 265(15):1985-1990. Available: <https://jamanetwork.com/journals/jama/article-abstract/385646>
4. Cole MP, Jones CTA, Todd IDH. A new anti-oestrogenic agent in late breast cancer: An early clinical appraisal of ICI46474. *British Journal of Cancer*. 1971; 25(2), 270.
5. Skouby SO, Pan K, Thompson JR, Komm BS, Mirkin S. Effects of conjugated estrogens/bazedoxifene on lipid and coagulation variables: A randomized placebo-and active-controlled trial. *Menopause*. 2015;22(6): 640-649. Available: https://journals.lww.com/menopausejournal/Abstract/2015/06000/Effects_of_conjugated_estrogens_bazedoxifene_on.10.aspx
6. Fabian CJ, Nye L, Powers KR, Nydegger JL, Kreutzjans AL, Phillips TA, Goodman ML. Effect of bazedoxifene and conjugated estrogen (Duavee) on breast cancer risk biomarkers in high-risk women: A pilot study. *Cancer Prevention Research*. 2019; 12(10):711-720. Available: <https://cancerpreventionresearch.aacrjournals.org/content/canprevres/12/10/711.full.pdf>
7. Kawate H, Takayanagi R. Efficacy and safety of bazedoxifene for postmenopausal osteoporosis. *Clinical Interventions in Aging*. 2011;6:151. Available: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3131985/>
8. Silverman SL, Christiansen C, Genant HK, Vukicevic S, Zanchetta JR, de Villiers TJ, Chines AA. Efficacy of bazedoxifene in reducing new vertebral fracture risk in postmenopausal women with osteoporosis: results from a 3-year, randomized, placebo-, and active-controlled clinical trial. *Journal of Bone and Mineral Research*. 2008;23(12):1923-1934. Available: <https://asbmr.onlinelibrary.wiley.com/doi/pdf/10.1359/jbmr.080710>
9. Kanis JA, Johansson H, Oden A, McCloskey EV. Bazedoxifene reduces vertebral and clinical fractures in postmenopausal women at high risk assessed with FRAX®. *Bone*. 2009;44(6): 1049-1054. Available: <https://www.sciencedirect.com/science/article/abs/pii/S8756328209004530>
10. Lindsay R, Gallagher JC, Kagan R, Pickar JH, Constantine G. Efficacy of tissue-selective estrogen complex of bazedoxifene/conjugated estrogens for osteoporosis prevention in at-risk postmenopausal women. *Fertility and Sterility*. 2009;92(3):1045-1052. Available: <https://www.sciencedirect.com/science/article/abs/pii/S0015028209013041>

11. Komm BS, Mirkin S, Jenkins SN. Development of conjugated estrogens/bazedoxifene, the first tissue selective estrogen complex (TSEC) for management of menopausal hot flashes and postmenopausal bone loss. *Steroids*. 2014;90:71-81. Available:<https://www.sciencedirect.com/science/article/abs/pii/S0039128X14001433>
12. Lobo RA, Pinkerton JV, Gass ML, Dorin MH, Ronkin S, Pickar JH, Constantine G. Evaluation of bazedoxifene/conjugated estrogens for the treatment of menopausal symptoms and effects on metabolic parameters and overall safety profile. *Fertility and Sterility*. 2009;92(3):1025-1038.
13. Stovall DW, Tanner-Kurtz K, Pinkerton JV. Tissue-selective estrogen complex bazedoxifene and conjugated estrogens for the treatment of menopausal vasomotor symptoms. *Drugs*. 2011;71(13):1649-1657.
14. Peng L, Luo Q, Lu H. Efficacy and safety of bazedoxifene in postmenopausal women with osteoporosis: A systematic review and meta-analysis. *Medicine*. 2017;96(49). Available:<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5728840/>
15. Bansal R, Aggarwal N. Menopausal hot flashes: a concise review. *Journal of Mid-life Health*. 2019;10(1):6. Available:<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6459071/>
16. Parish SJ, Gillespie JA. The evolving role of oral hormonal therapies and review of conjugated estrogens/bazedoxifene for the management of menopausal symptoms. *Postgraduate Medicine*. 2017;129(3):340-351. Available:<https://www.tandfonline.com/doi/abs/10.1080/00325481.2017.1281083>
17. Lewis-Wambi JS, Kim H, Curpan R, Grigg R, Sarker MA, Jordan VC. The selective estrogen receptor modulator bazedoxifene inhibits hormone-independent breast cancer cell growth and down-regulates estrogen receptor α and cyclin D1. *Molecular Pharmacology*. 2011;80(4):610-620.
18. Wardell SE, Nelson ER, Chao CA, McDonnell DP. Bazedoxifene exhibits antiestrogenic activity in animal models of tamoxifen-resistant breast cancer: implications for treatment of advanced disease. *Clinical Cancer Research*. 2013;19(9):2420-2431.
19. Wardell SE, Ellis MJ, Alley HM, Eisele K, VanArsdale T, Dann SG, Crowder R. Efficacy of SERD/SERM Hybrid-CDK4/6 inhibitor combinations in models of endocrine therapy-resistant breast cancer. *Clinical Cancer Research*. 2015;21(22):5121-5130. Available:<https://clincancerres.aacrjournals.org/content/clincanres/21/22/5121.full.pdf>
20. Fanning SW, Jeselsohn R, Dharmarajan V, Mayne CG, Karimi M, Buchwalter G, Lainé M. The SERM/SERD bazedoxifene disrupts ESR1 helix 12 to overcome acquired hormone resistance in breast cancer cells. *Elife*. 2018;7:37161. Available:<https://elifesciences.org/articles/37161>
21. Toy W, Shen Y, Won H, Green B, Sakr R A, Will M, Hudis C. ESR1 ligand-binding domain mutations in hormone-resistant breast cancer. *Nature Genetics*. 2013;45(12):1439. Available:<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3903423/>
22. Fu S, Chen X, Lo HW, Lin J. Combined bazedoxifene and paclitaxel treatments inhibit cell viability, cell migration, colony formation, and tumor growth and induce apoptosis in breast cancer. *Cancer Letters*. 2019;448:11-19.
23. Tian J, Chen X, Fu S, Zhang R, Pan L, Cao Y, Zhang Y. Bazedoxifene is a novel IL-6/GP130 inhibitor for treating triple-negative breast cancer. *Breast Cancer Research and Treatment*. 2019;175(3):553-566.
24. Boulanger MJ, Chow DC, Brevnova EE, Garcia KC. Hexameric structure and assembly of the interleukin-6/IL-6 α -receptor/gp130 complex. *Science*. 2003;300(5628):2101-2104.
25. Dethlefsen C, Højfeldt G, Hojman P. The role of intratumoral and systemic IL-6 in breast cancer. *Breast Cancer Research and Treatment*. 2013;138(3):657-664.
26. Kishimoto T, Akira S, Taga T. Interleukin-6 and its receptor: a paradigm for cytokines. *Science*. 1992;258(5082):593-597. Available:<https://science.sciencemag.org/content/258/5082/593.abstract>
27. Heo TH, Wahler J, Suh N. Potential therapeutic implications of IL-6/IL-

- 6R/gp130-targeting agents in breast cancer. *Oncotarget*. 2016;7(13): 15460.
Available: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4941253/>
28. Rabe B, Chalaris A, May U, Waetzig GH, Seegert D, Williams AS, Scheller J. Transgenic blockade of interleukin 6 transsignaling abrogates inflammation. *Blood, The Journal of the American Society of Hematology*. 2008;111(3):1021-1028.
Available: <https://ashpublications.org/blood/article/111/3/1021/25448/Transgenic-blockade-of-interleukin-6>
 29. Lust JA, Donovan KA, Kline MP, Greipp PR, Kyle RA, Maihle NJ. Isolation of an mRNA encoding a soluble form of the human interleukin-6 receptor. *Cytokine*. 1992;4(2):96-100.
Available: <https://www.sciencedirect.com/science/article/abs/pii/S104346669290043Q>
 30. Mülberg J, Schooltink H, Stoyan T, Günther M, Graeve L, Buse G, Rose-John, S. The soluble interleukin-6 receptor is generated by shedding. *European Journal of Immunology*. 1993;23(2):473-480.
Available: <https://onlinelibrary.wiley.com/doi/abs/10.1002/eji.1830230226>
 31. Available: <https://www.uniprot.org/uniprot/P08887> Downloaded 10/26/2020
 32. Kang S, Tanaka T, Narazaki M, Kishimoto T. Targeting interleukin-6 signaling in clinic. *Immunity*. 2019;50(4):1007-1023.
Available: <https://reader.elsevier.com/reader/sd/pii/S1074761319301438?token=46981F8181748D9822E435BA71F58C2E962365110E4B82A5E6ABFAA5083D067773B5BF7689F512D9B464F70C08B5FA3>
 33. Yawata H, Yasukawa K, Natsuka S, Murakami M, Yamasaki K, Hibi M, Taga T, Kishimoto T. Structure-function analysis of human IL-6 receptor: dissociation of amino acid residues required for IL-6-binding and for IL-6 signal transduction through gp130. *The EMBO Journal*. 1993;12(4): 1705–1712.
Available: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC413384/>
 34. Ni CW, Hsieh HJ, Chao YJ, Wang DL. Interleukin-6-induced JAK2/STAT3 signaling pathway in endothelial cells is suppressed by hemodynamic flow. *American Journal of Physiology-Cell Physiology*. 2004;287(3):771-780.
Available: <https://journals.physiology.org/doi/full/10.1152/ajpcell.00532.2003>
 35. Schust J, Sperl B, Hollis A, Mayer TU, Berg T. Stattic: A small-molecule inhibitor of STAT3 activation and dimerization. *Chemistry & Biology*. 2006;13(11):1235-1242.
Available: <https://www.sciencedirect.com/science/article/pii/S1074552106003784>
 36. Hirano T, Matsuda T, Nakajima K. Signal transduction through gp130 that is shared among the receptors for the interleukin 6 related cytokine subfamily. *Stem Cells*. 1994;12(3):262-277.
Available: <https://stemcells.journals.onlinelibrary.wiley.com/doi/abs/10.1002/stem.5530120303>
 37. Liu KD, Gaffen SL, Goldsmith MA. JAK/STAT signaling by cytokine receptors. *Current Opinion in Immunology*. 1998; 10(3):271-278.
Available: <https://www.sciencedirect.com/science/article/abs/pii/S0952791598801659>
 38. Calvisi DF, Ladu S, Gorden A, Farina M, Conner EA, Lee JS, Thorgeirsson SS. Ubiquitous activation of Ras and Jak/Stat pathways in human HCC. *Gastroenterology*. 2006;130(4):1117-1128.
Available: [https://www.gastrojournal.org/article/S0016-5085\(06\)00007-2/fulltext](https://www.gastrojournal.org/article/S0016-5085(06)00007-2/fulltext)
 39. Bromberg JF. Activation of STAT proteins and growth control. *Bioessays*. 2001;23(2): 161-169.
Available: [https://onlinelibrary.wiley.com/doi/abs/10.1002/1521-1878\(200102\)23:2%3C161::AID-BIES1023%3E3.0.CO;2-0](https://onlinelibrary.wiley.com/doi/abs/10.1002/1521-1878(200102)23:2%3C161::AID-BIES1023%3E3.0.CO;2-0)
 40. Johnson GL, Lapadat R. Mitogen-activated protein kinase pathways mediated by ERK, JNK, and p38 protein kinases. *Science*. 2002;298(5600):1911-1912.
Available: http://materiais.dbio.uevora.pt/B D/Crescimento/MAPK_Kinome.pdf
 41. Rawlings JS, Rosler KM, Harrison DA. The JAK/STAT signaling pathway. *Journal of Cell Science*. 2004;117(8):1281-1283.
Available: <https://jcs.biologists.org/content/joces/117/8/1281.full.pdf>
 42. Sansone P, Bromberg J. Targeting the interleukin-6/Jak/stat pathway in human malignancies. *Journal of Clinical Oncology*. 2012;30(9):1005.
Available: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3341105/>
 43. Calò V, Migliavacca M, Bazan V, Macaluso M, Buscemi M, Gebbia N, Russo A. STAT proteins: from normal control of

- cellular events to tumorigenesis. *Journal of Cellular Physiology*. 2003;197(2):157-168.
Available:<https://onlinelibrary.wiley.com/doi/pdf/10.1002/jcp.10364>
44. Chang Q, Daly L, Bromberg J. The IL-6 feed-forward loop: a driver of tumorigenesis. In *Seminars in immunology*. Academic Press. 2014;26(1):48-53.
Available:<https://www.sciencedirect.com/science/article/abs/pii/S1044532314000086>
 45. Wu X, Cao Y, Xiao H, Li C, Lin J. Bazedoxifene as a novel GP130 inhibitor for pancreatic cancer therapy. *Molecular Cancer Therapeutics*. 2016;15(11):2609-2619.
 46. Chen X, Tian J, Su GH, Lin J. Blocking IL-6/GP130 signaling inhibits cell viability/proliferation, glycolysis, and colony forming activity in human pancreatic cancer cells. *Current Cancer Drug Targets*. 2019;19(5):417-427.
Available:<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7032663/>
 47. Fu S, Lin J. Blocking interleukin-6 and interleukin-8 signaling inhibits cell viability, colony-forming activity, and cell migration in human triple-negative breast cancer and pancreatic cancer cells. *Anticancer Research*. 2018;38(11):6271-6279.
Available:<http://ar.iiarjournals.org/content/38/11/6271.short>
 48. Wu X, Xiao H, Li C, Lin J. Abstract A177: Repositioning bazedoxifene as a novel inhibitor of IL-6/GP130 signaling for pancreatic cancer therapy. In: *Proceedings of the AACR-NCI-EORTC International Conference: Molecular Targets and Cancer Therapeutics*. Boston, MA. Philadelphia (PA): AACR; *Mol Cancer Ther* Abstract nr A177. 2015;14(12Suppl 2):5-9.
Available:https://mct.aacrjournals.org/content/14/12_Supplement_2/A177.short
 49. Garbers C, Aparicio-Siegmund S, Rose-John S. The IL-6/gp130/STAT3 signaling axis: recent advances towards specific inhibition. *Current Opinion in Immunology*. 2015;34:75-82.
Available:<https://www.sciencedirect.com/science/article/abs/pii/S0952791515000370>
 50. Rosell R, Bertran-Alamillo J, Molina MA, Taron M. IL-6/gp130/STAT3 signaling axis in cancer and the presence of in-frame gp130 somatic deletions in inflammatory hepatocellular tumors; 2009.
Available:<https://www.futuremedicine.com/doi/abs/10.2217/fon.09.3>
 51. Geletu M, Arulanandam R, Chevalier S, Saez B, Larue L, Feracci H, Raptis L. Classical cadherins control survival through the gp130/Stat3 axis. *Biochimica et Biophysica Acta (BBA)-Molecular Cell Research*. 2013;1833(8):1947-1959.
Available:<https://www.sciencedirect.com/science/article/pii/S0167488913001158>
 52. Johnson DE, O'Keefe RA, Grandis JR. Targeting the IL-6/JAK/STAT3 signalling axis in cancer. *Nature Reviews Clinical Oncology*. 2018;15(4):234.
Available:<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5858971/>
 53. Taher MY, Davies DM, Maher J. The role of the interleukin (IL)-6/IL-6 receptor axis in cancer. *Biochemical Society Transactions*. 2018;46(6):1449-1462.
Available:<https://portlandpress.com/biochemsoctrans/article-abstract/46/6/1449/67638>
 54. Xiao H, Bid HK, Chen X, Wu X, Wei J, Bian Y, Lin J. Repositioning Bazedoxifene as a novel IL-6/GP130 signaling antagonist for human rhabdomyosarcoma therapy. *PLoS One*. 2017;12(7):0180297.
 55. Yadav A, Kumar B, Teknos TN, Kumar P. Bazedoxifene enhances the anti-tumor effects of cisplatin and radiation treatment by blocking IL-6 signaling in head and neck cancer. *Oncotarget*. 2017;8(40):66912.
 56. Tian J, Chen X, Fu S, Zhang R, Pan L, Cao Y, Zhang Y. Bazedoxifene is a novel IL-6/GP130 inhibitor for treating triple-negative breast cancer. *Breast Cancer Research and Treatment*. 2019;175(3):553-566.
 57. Thilakasiri P, Huynh J, Poh AR, Tan CW, Nero TL, Tran K, Buchert M.. Repurposing the selective estrogen receptor modulator bazedoxifene to suppress gastrointestinal cancer growth. *EMBO Molecular Medicine*. 2019;11(4).
 58. Wei J, Ma L, Lai YH, Zhang R, Li H, Li C, Lin J. Bazedoxifene as a novel GP130 inhibitor for Colon Cancer therapy. *Journal of Experimental & Clinical Cancer Research*. 2019;38(1):63.
 59. Burkhardt C, Bühler L, Tihy M, Morel P, Forni M. Bazedoxifene as a novel strategy for treatment of pancreatic and gastric adenocarcinoma. *Oncotarget*. 2019;10(34):3198.
Available:<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6516716/>

60. Chen X, Wei J, Li C, Pierson CR, Finlay JL, Lin J. Blocking interleukin-6 signaling inhibits cell viability/proliferation, glycolysis, and colony forming activity of human medulloblastoma cells. *International Journal of Oncology*. 2018;52(2):571-578. Available:<https://www.spandidos-publications.com/10.3892/ijo.2017.4211>
61. Wu X, Lin J. FDA-approved drug bazedoxifene as a novel inhibitor of IL-6 and IL-11/GP130 signaling for osteosarcoma therapy [abstract]. In: *Proceedings of the American Association for Cancer Research Annual Meeting 2018; 2018 Apr 14-18; Chicago, IL. Philadelphia (PA): AACR; Cancer Res. 2018;78(13 Suppl):5455.* Available:https://cancerres.aacrjournals.org/content/78/13_Supplement/5455.short
62. Ma H, Yan D, Wang Y, Shi W, Liu T, Zhao C, Zhai M. Bazedoxifene exhibits growth suppressive activity by Targeting IL-6/GP130/STAT3 Signaling in Hepatocellular Carcinoma. *Cancer Sci*. 2019;110:950-961. Available:<https://onlinelibrary.wiley.com/doi/full/10.1111/cas.13940>
63. Li S, Tian J, Zhang H, Zhou S, Wang X, Zhang L, Ji Z. Down-regulating IL-6/GP130 targets improved the anti-tumor effects of 5-fluorouracil in colon cancer. *Apoptosis*. 2018;23(5-6):356-374. Available:<https://link.springer.com/article/10.1007/s10495-018-1460-0>
64. Mohammed A, Sei S, Shoemakers R, Grubbs CJ. Combination of bazedoxifene and lapatinib profoundly inhibits estrogen receptor positive (ER+) and Negative (ER-) mammary tumorigenesis. In: *Proceedings of the Annual Meeting of the American Association for Cancer Research 2020; 2020 Apr 27-28 and Jun 22-24. Philadelphia (PA): AACR; Cancer Res 2020;80(16 Suppl):Abstract nr 1099.* Available:https://cancerres.aacrjournals.org/content/80/16_Supplement/1099.short
65. Jeselsohn R, Guo H, Rees R, Barry WT, Barlett CH, Tung NM, Winer EP. Results from the phase Ib/II clinical trial of bazedoxifene and palbociclib in hormone receptor positive metastatic breast cancer. *Cancer Res*. 2019;79(Suppl4):Abstractnr-PD1. Available:<https://sabcs18.posterview.com/nosl/p/PD1-05>
66. Wu X, Xiao H, Wang R, Liu L, Li C, Lin J. Persistent GP130/STAT3 signaling contributes to the resistance of doxorubicin, cisplatin, and MEK inhibitor in human rhabdomyosarcoma cells. *Current Cancer Drug Targets*. 2016;16(7):631-638. Available:<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5014400/>
67. Bossen J, Uliczka K, Steen L, Pfefferkorn R, Mai MMQ, Burkhardt L, Roeder T. An EGFR-induced Drosophila lung tumor model identifies alternative combination treatments. *Molecular Cancer Therapeutics*. 2019;18(9):1659-1668. Available:<https://mct.aacrjournals.org/content/18/9/1659.abstract>
68. Saucedo LJ, Edgar BA. Filling out the Hippo pathway. *Nature reviews Molecular Cell Biology*. 2007;8(8):613-621.
69. Morice S, Danieau G, Rédini F, Brounais-Le-Royer B, Verrecchia F. Hippo/YAP Signaling Pathway: A Promising Therapeutic Target in Bone Paediatric Cancers?. *Cancers*. 2020;12(3):645. Available:<https://doi.org/10.3390/cancers12030645>. Available:<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7139637/>
70. Fu W, Zhao P, Li H, Fu H, Liu X, Liu Y, Fu W. Bazedoxifene enhances paclitaxel efficacy to suppress glioblastoma via altering Hippo/YAP pathway. *Journal of Cancer*. 2020;11(3):657. Available:<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6959043/>
71. He C, Mao D, Hua G, Lv X, Chen X, Angeletti PC, Lambert PF. The Hippo/YAP pathway interacts with EGFR signaling and HPV oncoproteins to regulate cervical cancer progression. *EMBO Molecular Medicine*. 2015;7(11):1426-1449. Available:<https://www.embopress.org/doi/full/10.15252/emmm.201404976>
72. Zhang Y, Xie P, Wang X, Pan P, Wang Y, Zhang H, Zhou X. YAP promotes migration and invasion of human glioma cells. *Journal of Molecular Neuroscience*. 2018; 64(2):262-272. Available:<https://link.springer.com/article/10.1007/s12031-017-1018-6>
73. Huang J, Wu S, Barrera J, Matthews K, Pan D. The Hippo signaling pathway coordinately regulates cell proliferation and apoptosis by inactivating Yorkie, the Drosophila Homolog of YAP. *Cell*. 2005; 122(3):421-434.

- Available:<https://www.sciencedirect.com/science/article/pii/S0092867405005520>
74. Entrez gene; 2012.
Available:<https://www.ncbi.nlm.nih.gov/gene?Db=gene&Cmd=ShowDetailView&TermToSearch=316>
75. Chen S, Austin-Muttitt K, Zhang LH, Mullins JG, Lau AJ. *In vitro* and *In silico* analyses of the inhibition of human aldehyde oxidase by bazedoxifene, lasofoxifene, and structural analogues. *Journal of Pharmacology and Experimental Therapeutics*. 2019;371(1): 75-86.
Available:<https://core.ac.uk/download/pdf/227518203.pdf>
76. Zhang W, Chai W, Zhu Z, Li X. Aldehyde oxidase 1 promoted the occurrence and development of colorectal cancer by up-regulation of expression of CD133. *International Immunopharmacology*. 2020;85 :106618.
Available:<https://www.sciencedirect.com/science/article/pii/S1567576920310146>
77. Vantaku V, Putluri V, Bader DA, Maity S, Ma J, Arnold JM, Dubrulle J. Epigenetic loss of AOX1 expression via EZH2 leads to metabolic deregulations and promotes bladder cancer progression. *Oncogene*. 2019;1-21.
Available:<https://www.nature.com/articles/s41388-019-0902-7>
78. Hou R, Xu H, Li H. Effect of bazedoxifene on expressions of VEGF, VEGFR2, COX-2 and inflammatory factors in a rat endometriosis model. *Tropical Journal of Pharmaceutical Research*. 2019;18(9).
Available:https://www.tjpr.org/admin/12389900798187/2019_18_9_2.pdf
79. Dembo AG, Aledort ES, Greene GL. CE+BZA combination therapy decreases breast cancer outgrowth in part through activation of the AHR pathway [abstract]. In: *Proceedings of the Annual Meeting of the American Association for Cancer Research 2020*; 2020 Apr 27-28 and Jun 22-24. Philadelphia (PA): AACR; *Cancer Res*. 2020;80(16 Suppl):4374.
80. Sato S, Shirakawa H, Tomita S, Ohsaki Y, Haketa K, Tooi O, Komai M. Low-dose dioxins alter gene expression related to cholesterol biosynthesis, lipogenesis, and glucose metabolism through the aryl hydrocarbon receptor-mediated pathway in mouse liver. *Toxicology and Applied Pharmacology*. 2008;229(1):10-19.
Available:<https://pubmed.ncbi.nlm.nih.gov/18295293/>
81. Feng S, Cao Z, Wang X. Role of aryl hydrocarbon receptor in cancer. *Biochimica et Biophysica Acta (BBA)-Reviews on Cancer*, 2013;1836(2):197-210.
Available:<https://www.sciencedirect.com/science/article/abs/pii/S0304419X13000279>
82. Murray IA, Patterson AD, Perdew GH. Aryl hydrocarbon receptor ligands in cancer: Friend and foe. *Nature Reviews. Cancer*. 2014;14(12):801–814.
Available:<https://doi.org/10.1038/nrc3846>.
Available:<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4401080/>
83. Zachrewski T, Harris M, Biegel L, Morrison V, Merchant M, Safe S. 6-Methyl-1, 3, 8-trichlorodibenzofuran (MCDF) is an antiestrogen in human and rodent cancer cell lines: Evidence for the role of the Ah receptor. *Toxicology and Applied Pharmacology*. 1992;113(2):311-318.
Available:<https://pubmed.ncbi.nlm.nih.gov/1313996/>
84. Perry R, Thompson CA, Earnhardt JN, Wright DJ, Bailey S, Komm B, Cukierski MA. Renal tumors in male rats following long-term administration of bazedoxifene, a tissue-selective estrogen receptor modulator. *Toxicologic pathology*. 2013; 41(7):1001-1010.
85. Friedrich A, Olejniczak K. Evaluation of carcinogenicity studies of medicinal products for human use authorised via the European centralised procedure (1995–2009). *Regulatory Toxicology and Pharmacology*. 2011;60(2):225-248.
86. Gudbrandsdottir G, Aarstad HH, Bostad L, Hjelle KM, Aarstad HJ, Bruserud Ø, Beisland C. Serum levels of the IL-6 family of cytokines predict prognosis in renal cell carcinoma (RCC). *Cancer Immunology, Immunotherapy*. 2020;1-12.
Available:<https://link.springer.com/article/10.1007/s00262-020-02655-z>
87. Michalaki V, Syrigos K, Charles P, Waxman J. Serum levels of IL-6 and TNF- α correlate with clinicopathological features and patient survival in patients with prostate cancer. *British Journal of Cancer*. 2004;90(12):2312-2316.
Available:<https://www.nature.com/articles/6601814>
88. Nakashima J, Tachibana M, Horiguchi Y, Oya M, Ohigashi T, Asakura H, Murai M. Serum interleukin 6 as a prognostic factor

- in patients with prostate cancer. *Clinical Cancer Research*. 2000;6(7):2702-2706. Available: <https://clincancerres.aacrjournals.org/content/6/7/2702>
89. Drachenberg DE, Elgamal AAA, Rowbotham R, Peterson M, Murphy GP. Circulating levels of interleukin-6 in patients with hormone refractory prostate cancer. *The Prostate*. 1999;41(2): 127-133. Available: [https://onlinelibrary.wiley.com/doi/abs/10.1002/\(SICI\)1097-0045\(19991001\)41:2%3C127::AID-PROS7%3E3.0.CO;2-H](https://onlinelibrary.wiley.com/doi/abs/10.1002/(SICI)1097-0045(19991001)41:2%3C127::AID-PROS7%3E3.0.CO;2-H)
 90. Chung YC, Chang YF. Serum interleukin-6 levels reflect the disease status of colorectal cancer. *Journal of Surgical Oncology*. 2003;83(4):222-226. Available: <https://onlinelibrary.wiley.com/doi/abs/10.1002/jso.10269>
 91. Nikiteas NI, Tzanakis N, Gazouli M, Rallis G, Daniilidis K, Theodoropoulos G, Peros G. Serum IL-6, TNF α and CRP levels in Greek colorectal cancer patients: prognostic implications. *World Journal of Gastroenterology: WJG*. 2005;11(11): 1639. Available: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4305945/>
 92. Ueda T, Shimada E, Urakawa, T. Serum levels of cytokines in patients with colorectal cancer: possible involvement of interleukin-6 and interleukin-8 in hematogenous metastasis. *Journal of Gastroenterology*. 1994;29(4):423-429. Available: <https://link.springer.com/article/10.1007%2FBF02361238>
 93. Yanagawa H, Sone S, Takahashi Y, Haku T, Yano S, Shinohara T, Ogura T. Serum levels of interleukin 6 in patients with lung cancer. *British Journal of Cancer*. 1995; 71(5):1095-1098. Available: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2033769/pdf/brjcancer00051-0199.pdf>
 94. Knüpfer H, Preiß R. Significance of interleukin-6 (IL-6) in breast cancer. *Breast cancer research and treatment*, 2007; 102(2):129-135. Available: <https://link.springer.com/article/10.1007/s10549-006-9328-3>
 95. Ikeguchi M, Hatada T, Yamamoto M, Miyake T, Matsunaga T, Fukumoto Y, Tatebe S. Serum interleukin-6 and-10 levels in patients with gastric cancer. *Gastric Cancer*. 2009;12(2):95-100. Available: <https://link.springer.com/article/10.1007/s10120-009-0509-8>
 96. Sanguinete MMM, Oliveira PHD, Martins-Filho A, Micheli DC, Tavares-Murta BM, Murta EFC, Nomelini RS. Serum IL-6 and IL-8 correlate with prognostic factors in ovarian cancer. *Immunological investigations*. 2017;46(7):677-688. Available: <https://www.tandfonline.com/doi/abs/10.1080/08820139.2017.1360342>
 97. Duffy SA, Taylor JM, Terrell JE, Islam M, Li Y, Fowler KE, Teknos TN. Interleukin-6 predicts recurrence and survival among head and neck cancer patients. *Cancer: Interdisciplinary International Journal of the American Cancer Society*. 2008;113(4): 750-757. Available: <https://acsjournals.onlinelibrary.wiley.com/doi/full/10.1002/cncr.23615>
 98. Heikkilä K, Ebrahim S, Lawlor DA. Systematic review of the association between circulating interleukin-6 (IL-6) and cancer. *European Journal of Cancer*. 2008;44(7):937-945. Available: <https://www.sciencedirect.com/science/article/abs/pii/S0959804908001937>
 99. Klausen T, Olsen NV, Poulsen TD, Richalet JP, Pedersen BK. Hypoxemia increases serum interleukin-6 in humans. *European Journal of Applied Physiology and Occupational Physiology*. 1997;76(5):480-482. Available: <https://link.springer.com/article/10.1007/s004210050278>
 100. De Schutter H, Landuyt W, Verbeken E, Goethals L, Hermans R, Nuyts S. The prognostic value of the hypoxia markers CA IX and GLUT 1 and the cytokines VEGF and IL 6 in head and neck squamous cell carcinoma treated by radiotherapy±chemotherapy. *BMC Cancer*. 2005;5(1):42. Available: <https://link.springer.com/article/10.1186/1471-2407-5-42>
 101. Jiang J, Wang GZ, Wang Y, Huang HZ, Li WT, Qu XD. Hypoxia-induced HMGB1 expression of HCC promotes tumor invasiveness and metastasis via regulating macrophage-derived IL-6. *Experimental Cell Research*. 2018;367(1):81-88. Available: <https://www.sciencedirect.com/science/article/abs/pii/S0014482718301745>
 102. Xu K, Zhan Y, Yuan Z, Qiu Y, Wang H, Fan G, Zhang J. Hypoxia induces drug resistance in colorectal cancer through the HIF-1 α /miR-338-5p/IL-6 feedback loop. *Molecular Therapy*. 2019;27(10): 1810-

1824.
Available:<https://www.sciencedirect.com/science/article/pii/S1525001619302643>
103. Weng YS, Tseng HY, Chen YA, Shen PC, Al Haq AT, Chen LM, Hsu HL. MCT-1/miR-34a/IL-6/IL-6R signaling axis promotes EMT progression, cancer stemness and M2 macrophage polarization in triple-negative breast cancer. *Molecular cancer*, 2019;18(1):1-15.
Available:<https://molecularcancer.biomedcentral.com/articles/10.1186/s12943-019-0988-0>
 104. Wu X, Tao P, Zhou Q, Li Jm, Yu Z, Wang X, Liu B. IL-6 secreted by cancer-associated fibroblasts promotes epithelial-mesenchymal transition and metastasis of gastric cancer via JAK2/STAT3 signaling pathway. *Oncotarget*. 2017;8(13):20741.
Available:<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5400541/>
 105. Chan LC, Li CW, Xia W, Hsu JM, Lee HH, Cha JH, Zha Z. IL-6/JAK1 pathway drives PD-L1 Y112 phosphorylation to promote cancer immune evasion. *The Journal of Clinical Investigation*. 2019;129(8).
Available:<https://www.jci.org/articles/view/126022>
 106. Ferraresi A, Phadngam S, Morani F, Galetto A, Alabiso O, Chiorino G, Isidoro C. Resveratrol inhibits IL-6-induced ovarian cancer cell migration through epigenetic up-regulation of autophagy. *Molecular Carcinogenesis*. 2017;56(3):1164-1181.
Available:<https://onlinelibrary.wiley.com/doi/abs/10.1002/mc.22582>
 107. Wang L, Cao L, Wang H, Liu B, Zhang Q, Meng Z, Xu K. Cancer-associated fibroblasts enhance metastatic potential of lung cancer cells through IL-6/STAT3 signaling pathway. *Oncotarget*. 2017;8(44):76116.
Available:<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5652691/>
 108. He JY, Wei XH, Li SJ, Liu Y, Hu HL, Li ZZ., Sun L. Adipocyte-derived IL-6 and leptin promote breast Cancer metastasis via upregulation of Lysyl Hydroxylase-2 expression. *Cell Communication and Signaling*, 2018;16(1):1-19.
Available:<https://biosignaling.biomedcentral.com/articles/10.1186/s12964-018-0309-z>
 109. Bharti R, Dey G, Mandal M. Cancer development, chemoresistance, epithelial to mesenchymal transition and stem cells: A snapshot of IL-6 mediated involvement. *Cancer Letters*. 2016;375(1):51-61.
Available:<https://www.sciencedirect.com/science/article/abs/pii/S0304383516301264>
 110. Yousefi H, Momeny M, Ghaffari SH, Parsanejad N, Poursheikhani A, Javadikooshesh S, Sankanian G. IL-6/IL-6R pathway is a therapeutic target in chemoresistant ovarian cancer. *Tumori Journal*. 2019;105(1):84-91.
Available:<https://journals.sagepub.com/doi/full/10.1177/0300891618784790>
 111. Xiong S, Wang R, Chen Q, Luo J, Wang J, Zhao Z, Cheng B. Cancer-associated fibroblasts promote stem cell-like properties of hepatocellular carcinoma cells through IL-6/STAT3/Notch signaling. *American Journal of Cancer Research*. 2018;8(2):302.
Available:<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5835697/>
 112. Wang T, Song P, Zhong T, Wang X, Xiang X, Liu Q, Hu Y. The inflammatory cytokine IL-6 induces FRA1 deacetylation promoting colorectal cancer stem-like properties. *Oncogene*. 2019;38(25):4932-4947.
Available:<https://www.nature.com/articles/s41388-019-0763-0>
 113. Qin X, Yan M, Wang X, Xu Q, Wang X, Zhu X, Chen W. Cancer-associated fibroblast-derived IL-6 promotes head and neck cancer progression via the osteopontin-NF-kappa B signaling pathway. *Theranostics*. 2018;8(4):921.
Available:<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5817102/>
 114. Cui G, Yuan A, Sun, Z, Zheng W, Pang Z. IL-1 β /IL-6 network in the tumor microenvironment of human colorectal cancer. *Pathology-Research and Practice*. 2018;214(7):986-992.
Available:<https://www.sciencedirect.com/science/article/abs/pii/S0344033818300633>
 115. Wolfe AR, Trenton NJ, Debeb BG, Larson R, Ruffell B, Chu K, Woodward WA. Mesenchymal stem cells and macrophages interact through IL-6 to promote inflammatory breast cancer in pre-clinical models. *Oncotarget*. 2016;7(50):82482.
Available:<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5347707/>
 116. Hashizume M, Hayakawa N, Suzuki M, Mihara M. IL-6/sIL-6R trans-signaling, but not TNF- α induced angiogenesis in a HUVEC and synovial cell co-culture

- system. *Rheumatology International*. 2009; 29(12):1449-1454.
Available:<https://link.springer.com/article/10.1007%2Fs00296-009-0885-8>
117. Huang SP, Wu MS, Shun CT, Wang HP, Lin MT, Kuo ML, Lin JT. Interleukin-6 increases vascular endothelial growth factor and angiogenesis in gastric carcinoma. *Journal of Biomedical Science*. 2004;11(4):517-527.
Available:<https://www.karger.com/Article/Abstract/77902>
 118. Liu Q, Li G, Li R, He Q, Deng L, Zhang C, Zhang J. IL-6 promotion of glioblastoma cell invasion and angiogenesis in U251 and T98G cell lines. *Journal of Neuro-oncology*, 2010;100(2):165-176.
Available:<https://link.springer.com/article/10.1007%2Fs11060-010-0158-0>
 119. Fan Y, Ye J, Shen F, Zhu Y, Yeghiazarians Y, Zhu W, Yang GY. Interleukin-6 stimulates circulating blood-derived endothelial progenitor cell angiogenesis *In vitro*. *Journal of Cerebral Blood Flow & Metabolism*.2008;28(1):90-98.
Available:<https://journals.sagepub.com/doi/full/10.1038/sj.jcbfm.9600509>
 120. Jee SH, Chu CY, Chiu HC, Huang YL, Tsai WL, Liao YH, Kuo ML. Interleukin-6 induced basic fibroblast growth factor-dependent angiogenesis in basal cell carcinoma cell line via JAK/STAT3 and PI3-kinase/Akt pathways. *Journal of Investigative Dermatology*. 2004;123(6): 1169-1175.
Available:<https://www.sciencedirect.com/science/article/pii/S0022202X15320546>
 121. Pettersen K, Andersen S, Degen S, Tadini V, Grosjean J, Hatakeyama S, Romundstad PR. Cancer cachexia associates with a systemic autophagy-inducing activity mimicked by cancer cell-derived IL-6 trans-signaling. *Scientific Reports*. 2017;7(1):1-16.
Available:<https://www.nature.com/articles/s41598-017-02088-2>
 122. Strassmann G, Fong M, Kenney JS, Jacob CO. Evidence for the involvement of interleukin 6 in experimental cancer cachexia. *The Journal of Clinical Investigation*. 1992;89(5)1681-1684.
Available:<https://dm5migu4zj3pb.cloudfront.net/manuscripts/115000/115767/JCI92115767.pdf>
 123. Narsale AA, Carson JA. Role of IL-6 in cachexia—therapeutic implications. *Current Opinion in Supportive and Palliative Care*. 2014;8(4):321.
Available:<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4323347/>
 124. Wu L, Deng Z, Peng Y, Han L, Liu J, Wang L, Wei H. Ascites-derived IL-6 and IL-10 synergistically expand CD14+ HLA-DR-low myeloid-derived suppressor cells in ovarian cancer patients. *Oncotarget*. 2017; 8(44):76843.
Available:<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5652747/>
 125. Meng J, Zhang XT, Liu XL, Fan L, Li C, Sun Y, Zhang T. WSTF promotes proliferation and invasion of lung cancer cells by inducing EMT via PI3K/Akt and IL-6/STAT3 signaling pathways. *Cellular Signalling*. 2016;28(11):1673-1682.
Available:<https://www.sciencedirect.com/science/article/abs/pii/S0161589017301906>
 126. Duggan ST, McKeage K. Bazedoxifene. *Drugs*. 2011;71(16):2193-2212.
Available:<https://link.springer.com/article/10.2165/11207420-000000000-00000>
 127. Adami S, Palacios S, Rizzoli R, Levine AB, Sutradhar S, Chines AA. The efficacy and safety of bazedoxifene in postmenopausal women by baseline kidney function status. *Climacteric*. 2014; 17(3):273-284.
Available:<https://www.tandfonline.com/doi/abs/10.3109/13697137.2013.830605>
 128. McKeand W, Baird-Bellaire S, Ermer J, Patat A. Pharmacokinetics and safety of bazedoxifene in hepatically impaired and healthy postmenopausal women. *Clinical Pharmacology in Drug Development*. 2018; 7(4):365-372.
Available:<https://accp1.onlinelibrary.wiley.com/doi/abs/10.1002/cpdd.438>
 129. Palacios S. Efficacy and safety of bazedoxifene, a novel selective estrogen receptor modulator for the prevention and treatment of postmenopausal osteoporosis. *Current Medical Research and Opinion*. 2010;26(7):1553-1563.
Available:<https://www.tandfonline.com/doi/abs/10.1185/03007991003795873>
 130. Millán MM, Castañeda S. Sex Hormones and Related Compounds, Including Hormonal Contraceptives. In *Side Effects of Drugs Annual*. Elsevier. 2015;37:499-511.
Available:<https://www.sciencedirect.com/science/article/pii/S0378608015000239>

131. Lušin TT, Tomašić T, Trontelj J, Mrhar A, Peterlin-Mašič L. *In vitro* bioactivation of bazedoxifene and 2-(4-hydroxyphenyl)-3-methyl-1H-indol-5-ol in human liver microsomes. *Chemico-biological Interactions*. 2012;197(1):8-15. Available:<https://www.sciencedirect.com/science/article/abs/pii/S0009279712000312>

© 2020 Koltai; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
<http://www.sdiarticle4.com/review-history/62890>