

International Journal of Plant & Soil Science

Volume 36, Issue 3, Page 298-305, 2024; Article no.IJPSS.112803 ISSN: 2320-7035

# Plant Growth Promoting Rhizobacteria for Sustainable Production of Sugarcane and Rice

# P. P. Khandagale <sup>a++\*</sup>, S. S. Kansara <sup>a#</sup>, Jay Padsala <sup>a++</sup> and P. R. Patel <sup>b†</sup>

<sup>a</sup> Department of Plant Pathology, N. M. College of Agriculture, NAU, Navsari, India <sup>b</sup> Department of Plant Protection, ASPEE College of Horticulture, NAU, Navsari, India.

#### Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

#### Article Information

DOI: 10.9734/IJPSS/2024/v36i34427

#### **Open Peer Review History:**

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/112803

**Review Article** 

Received: 04/12/2023 Accepted: 09/02/2024 Published: 15/02/2024

#### ABSTRACT

Sugarcane (*Sacchraum officinarum* L.) and rice (*Oryza sativa* L.) are important cash and staple crop in the world respectively. In today's population explosion the lots of pressure on cropping land to mitigate the feed need of consumers with judicious use of chemical fertilizers without considering the health of soil and its sustainability. The rhizobacteria live region around roots (2-80 mm) of crop with divers, dynamic and complex microflora having capacity with direct and indirect beneficial effects on crop health by availability of different nutrient, siderophores, rhizo deposits, auxins and role with bioremediation. There are different types of bacteria is isolated from rhizosphere of sugarcane and rice having Bacillus, Rhizobium, comamonas, cyanaobacteria, Nodosilinea, Levinella and Pseudomonas genera with most efficient nitrogen producer, solubilizers of inorganic

++ Ph.D. Research Scholar;

- # Assistant Professor;
- <sup>†</sup>Associate Professor and Head;

\*Corresponding author: E-mail: p2khandagale@gmail.com;

Int. J. Plant Soil Sci., vol. 36, no. 3, pp. 298-305, 2024

phosphate, potash and other macro and micronutrient. In rice microbiome as Levinella, Psudomonas Anaeromyxobacter, Arenimonas, Arthrobacter, Bacillus, Bellilinea Proteobacteria, Chloroflexi, Actinobacteria, Acidobacteria and cyanobacteria with strains of Nitrosomonas and Nitrobacter are further divided in vegetative and reproductive growth stage of rice crop. The article give brief idea about the large arena of them.

Keywords: Rhizobacteria; sugarcane; rice; sustainable production.

#### 1. INTRODUCTION

Rhizosphere give the term by Hiltner (1904) as narrow soil region near root zone having different secretion and associated soil microorganism microbiome root as communication .They feed on sloughed off plant cells called rhizodeposition and vice versa root released protein called root exudates. This symbiosis influence plant growth providing space to produce some allochemicals to control rhizospheric neiahbors. Having the effect enhance the redox potential, ionic balance, pressure with formation osmatic of soil aggregates and prevent the crop roots from desiccation in abiotic stress Sugarcane (Sacchraum officinarum L.) and rice (Oryza sativa L.) are important cash and staple crop in the world respectively. Sugarcane gives us tea sugar, brown sugar, alcohol, spirit, bagasse, press mud cakes, molasses with renewable energy sources for community transport with methanol blending in petrol. Every sugar factory enhances the socio economic status of the producing farmer in their locality with direct indirect employments opportunity generated by them. Now a days bio electricity generated by using the sugarcane wastes. For its sustainable production with soil management required use of different biofertilizers with bacterial inoculations consortia. "In sugarcane various kinds of bacterial endophytes such as Glucano acetobacter. Psudomonas. Azospirillum, Burkholderia. Raoultella. Ralstonia Mesorhizobium, Ochrobactrum, Sphingomonas, Novosphingobium, Pantoea and Bacillus were found and they are perform various role like promote growth, biological nitrogen fixation, production of phyto stimulants like - IAA, GA, Ethvlene and cytokinin, phosphate mineralization, potash solubilization, iron carrier formation, saponin and glycosidase production, production of 1-amino-cyclopropane-1-carboxylic deaminase as the antibiotics acid and development of ISR and SAR throughout its life period sustain the crop in biotic and abiotic stress environmental condition" .

"Rice the main staple food for about half of the world's population, microorganisms exert positive influences on plant growth, yield potential production, and health of rice. A variety of microbial species as actinomycetes and Grampositive bacteria reside in the rhizosphere and the phyllosphere of plants and also have multiple roles as symbiotic endophytes. They alter the morphology of host plants, enhance their growth. health, and vield, and reduce their vulnerability abiotic to biotic and stresses. Bacterial communities are usually dominated by the Anaeromyxobacter, Arenimonas, Arthrobacter, B acillus, Bellilinea Proteobacteria, Chloroflexi. and Actinobacteria, Acidobacteria, and cyanobacteria which help in uptake of nitrogen and other growth promoting role in rice" [1].

### 2. SUGARCANE

#### 2.1 *Azosprillum* and sugarcane

"Azospirillum is facultative endophytic, polymorphic, obligate can grow under low PO2 soil condition . Are microgram-negative aerophilic, rods and often associated with interior and surfaces roots of cereals and grasses" [2]. "It is very well known for its nitrogen-fixing ability and can fix at least 10 mg N g1 of carbohydrate and higher production of indole acetic acid. Three species of this genus namely. Azospirillum amazonense, A. brasilense, and A. lipoferum have been isolated .Microbial from sugarcane roots consortia containing the species. Azospirillum brasilense, and Bacillus subtilis have shown great potential to cycle nutrients from crop residues and restore soil fertility. The seed treatment enhances the root volume at 86% and 100% increase than untreated ones.The inoculation with Azosprillum brasilense + B. subtilis associated with 45 ha<sup>-1</sup> of kq P<sub>2</sub>O<sub>5</sub> also increases sugar yield, resulting in a savings of 75% of the recommended P<sub>2</sub>O<sub>5</sub> rate" [2].

# 2.2 *Enterobacteriaceae* Family Bacteria and Sugarcane

"The members of Enterobacteriaceae a large family of bacteria are rod-shaped, gram-negative, facultative anaerobes, motile or non-motile and most of them reduce nitrate to nitrite the dominating nitrogen-fixing bacteria isolated from sugarcane rice, wheat, sorghum, grasses, and dicotyledonous plants roots. The three genera the Enterobacteriaceae belona to as Enterobacter cloacae and Klebsiella pneumoniae and Pantoea were isolated from all parts of the sugarcane. Three species of this genus, Pantoea ananatis, P. herbicola, and P. stewartii have been isolated from roots, stem, and leaves of Brazilian sugarcane plants" [3] capable of nitrogen fixation.

#### 2.3 Gluconacetobacter and sugarcane

"This belonas aenus to the familv Acetobacteraceae to produce acetic acid and usually acid-tolerant and nitrogen fixing and grow well below pH 5.0. They are gram-negative, aerobic, and rod-shaped bacteria. This organism has lack of nitrate reductase and only partial inhibition of nitrogenase activity by ammonium ions, enables it to fix nitrogen in the presence of soil nitrogenease is also a phytohormone producer. In sugarcane endophytic Gluconacetobacter diazotrophicus and its abilities to fix nitrogen and disease resistance studied"[4]. dizotrophicus G. (leaf inhibit Xanthomonas albilineans scald disease) [5] .At Central Sugarcane Research Padegaon, Dist .Satara Station (Maharashtra)studied the effect of Glucanoacetobacter diazotrophicus @ 10kg/ha (Set treatment) with other six treatment was from year 2014-2018 on preseason carried sugarcane on CoM 0265 variety with (75% P<sub>2</sub>O<sub>5</sub>, 100% K<sub>2</sub>O + Recommended FYM (Suru-20 t/ha, Preseason- 25 t/ha) and PSB application common to all treatments except T1 RDF, found that treatment with 50% N+ Glucanoacetobacter set treatment recorded highest tillering count at 120 DAP (167.05 thousands/ha) and no. of mill liable cane .For cane yield same treatment recorded highest cane vield (57.54 t/ha).,CCS%,CCS yield and sucrose %.[6] .Same treand found in suru(Dec-Jan. Plantation) sugarcane as treatments with ( 50% N+ Glucanoacetobacter set treatment )recorded highest (162.00) tillering count at 120 DAP,NMC (53.31 thousand /ha) and cane yield. (52.17 t/ha. As regards CCS yield highest (7.98 t/ha), highest

purity (97.90), sucrose %(21.20) found with same trends. The endophytic bacterial count found in same tretment as in root (7.10x104, 11.20x104 and7.58x104), juice (16.53x104,29.63x104 and 18.96x104) and leaves (14.48x104, 30.01x104 and 16.35x104) at 4 months, 8 months and harvest stage of the crop, respectively. Having beneficial impact on sugar revcovery, NMC and other yield attributing parameters. Some scientist found that a new role for *G. diazotrophicus* induce systemic resistance against *Xanthomonas albilineans*-cause leaf scald disease of sugarcane [7].

### 2.4 Pseudomonas spp. and Sugarcane

"The belonas aenus to the familv Pseudomonadaceae with gram negative, rod shaped bacteria secreted soluble greenish fluorescent pigment called fluorescein in low availability of iron are facultative anaerobic require  $NO_3$  without  $O_2$  and some with  $O_2$  with multiple polar flagella as thev are chemolithotrophs. Haning large number of species with very well-known PGPR bacteria due its ability to produce phytohormones, to phosphate siderophores, antibiotics. production of antifungal solubilization and compounds. Some species also fix nitrogen in addition to above-mentioned characteristics. Pseudomonas fluorescens. Pseudomonas putida for PGPR and Pseudomonas aurantiaca, Pseudomonas chlororaphis for biocontrol agents in foliar spray due to the production of antifungal phenazine compounds. Pseudomonas spp. have been isolated from stem, root, leaves, and rhizosphere of sugarcane growing in Australia, Brazil, India, Pakistan, and South Africa. They are among most efficient solubilizers of inorganic phosphate" [8]. They produces phytohormones like indoleacetic acid, gibberellins, and cytokinin, siderophores, phosphateiron-sequestering solubilizing enzymes, and 1-aminocyclopropane-1-carboxylate (ACC) deaminase [9]. Growth hormones produced by the bacteria enhance the development of lateral roots and improve the plant's nutrient uptake from the rhizosphere.

# 2.5 PGPR and Biocontrol agents in Sugarcane

PGPR induce resistance in plants against fungal, bacterial, viral diseases and insects and nematodes attacks. PGPR bring about Induced resistance through fortifying the physical and mechanical strength of the cell wall as well as changing the physiological and biochemical reaction of the host leading to the synthesis of defense chemicals against the challenge pathogen. PGPR provide different mechanisms for suppressing plant pathogens having pre and post biochemical and cellular modification to suppress the invasion of disease pathogen. They include competition for nutrients and space, antibiosis by producing antibiotics viz pyrrolnitrin, pyocyanin, 2,4-diacetyl phloroglucinol and production of siderophores, viz., Pseudobactin which limits the availability iron necessary for the growth of of pathogens.

"The P. fluorescens strains from sugarcane reported antifungal activity against Fusarium oxysporum and Rhizoctonia bataticola" [10]. "The Ρ. putida. В. subtilis, strains and Stenotrophomonas maltophilia found control against local strains of red rot -Colletotrichum falcatum" [11] [12] checked the antifungal activity of Burkholderia isolates from the sugarcane rhizosphere, against U. scitaminea (sugarcane smut) and Fusarium spp. (stalk rot) [13].

## 3. RICE

In rice different growth stages with availability of water the PGPR having different species and its population. As vegetative stage bacterial population are differ from their reproductive stage. The ponding standing water in rice field rich in of Gram-negative bacteria plus algae. while in percolating water having only Gramnegative bacteria dominated by the Anaeromyxobacter, Arenimonas Arthrobacter, B acillus, Bellilinea, Proteobacteria, Chloroflexi. Actinobacteria, and Acidobacteria, and cyanobacteria.

### 3.1 Rice and Pantoea inoculant

"Bacteria belonging to the genus Pantoea are gram negative, highly diverse, aerobic or facultative anaerobic with peritrichous flagella in Enterobacteriaceae family though they are infecting rice with diseases some strains are till beneficial (P. ananatis, P. agglomerans, P .rodosii) showing a promising capacity to mediate the negative effects of arsenic stress as bioremediation and also act as phyto stimulants . They act as potential P solubilizes, N fixer ,IAA and salt tolerance with siderophore production rice endophyte Pantoea capacity. The agglomerans (YS19) showed nitrogen-fixing activity in vitro produced four different phytohormones, including IAA and promoted plant growth [14] "They found ubiquitous in roots and reduced at the time of tillering with subsequent stages" [15].

## 3.2 Rice and Bacillus spp.

Microorganisms in the endospheric Bacillus are aerobic, gram positive, soil bacterium affect the root morphology, tiller biomass when was applied as an inoculant biofertilizer. They synergists affects on chlorophyll content, plant height, number of tillers, tiller biomass and yield attributes in rice. The combination of TUAT-1 [13] (Bacillus pumilus strain) and 100% N (farmer recommended rate of N) resulted in the greatest tiller number and biomass at the maximum tillering stage, and positively affected other growth attributes and yield (4.89 ton ha-1) and TUAT-1 concluded that promoted root development which increased nutrient uptake from the soil by promoting the growth and development of roots.

The strain Bacillus subtilis sub sp. megatherium acts as the PGPR and bio control agent in rice so that plant can overcome the negative effect of environmental stress. Plant increased micronutrients uptake affect Phyto hormone homeostasis, stabilized membrane integrity, decreased leaf transpiration ,increased nutrition and metabolic activity with maintaining cell leaves turaor of rice under drought stress because of production of hormone IAA [16,17].

# 3.3 Rice and Siderophore-producing bacteria

"The genera Sphingomonas, Pseudomonas, Pantoea , Burkholderia and Enterobacter are primarily detected in rice plant tissues during plants' vegetative stage acts as siderophores, giving them greater access to iron and other elements in the soil. The Psudomonas fluroescens at field application enhanced the number of tillers, plant height and biomass with yield increase at 20-26 % over control" [18]. Inoculation with Beijerinckia indica in field increased no. of tillers and plant height with 25-29% increased yield [19].

### 3.4 Rice and Azospirilum Inoculant

They are surface colonizing, gram negative, free living, nitrogen fixing, polar flagellar moment forming cyst in unfavorable climatic condition. The inoculation of rice seedlings with *Azospirillum brasilense* promoted early tillering and the better reproductive performance of rice plants. It was found to significantly increase the grain-filling rate and the grain weight per plant at harvest time. [20] found that *Azospirillum* spp.(B510) strain in field condition enhanced no. of tillers and plant height with increased in yield at 17-25%.

#### 3.5 Rice and Psudomoans Inoculation

"In rice the inoculation with P. *dispersa* significantly improves the morphobiochemical characteristics of the plants such as increased enzymatic activities and reduced arsenic uptake into rice tissues" [21]. A study by, indicates that "Pseudomonas [22]and [23] fluorescens produces а volatile organic compound (pyrazine) that can suppress infection and physiological damage can give by Magnoporthe oryzae in rice".

# 3.6 Rice and *Rhizobium* bacterial inoculant

Though rice plants are not leguminous. So benefits of nitrogen not from N-fixation in nodules formed on plant roots because they can't have the Svm gene for endophytic colonization on rice root. However they having beneficial endophytic association [24,25] with Rhizobium different strains. They enhance crop health, yield and reduced the harmful effect of fungicides ,pesticides and synthetic chemicals. "In Egypt, it was found that the association of Rhizobium leguminosarum with rice plants could significantly increase the shoot and root growth, grain yield and nitrogen-use efficiency" [26] . "The growth responses of rice plants and vield to inoculation with two bacterial strains (R. (E11) and Rhizobium sp. leguminosarum (IRBG74) showed significantly increased grain and higher straw yields at the maturity stage" [27]. A multinational collaborative study with three field experiments in Egypt reported that the Rhizobium inoculation of rice plants significantly increased their biomass, nutrient uptake (due to more robust root architecture), grain yield, fertilizer efficiency, harvest index, and grain nutritional value. Further experiments of selected rhizobiaum endophytes on rice showed that they produced cell-bound cellulase and poly galacturonase enzymes that hydrolyze glycosidic bonds in plant cell walls plus certain bacteriocins that can inhibit the growth of undesirable microbes.

#### 3.7 Rice and PGPR as Disease Control

"Antifungal compounds produced bv B. subtilis show a strong synergistic inhibitory effect on the hyphal growth of *Pyricularia grisea* and *R*. solani the as causative agents for blast and sheath blight in rice" [28] . "Another study that employed а strain of endophytic bacterium Bacillus sp. (EBPBS4) and Serratia macrcescens used for managing sheath blight as antagonistic toward the fungal pathogen R. solani by decreasing its severity and incidences" [29-32]. Bacillus oryzicola reduced the bakanae disease (Gibberella fujikuroi) severity by 46-78% in field condition after its inoculation. The Pseudomonas spp. as biocontrol agents for resisting various soil-borne plant diseases [28] inhibits the mycelial growth of the sheath blight fungus R. solani by increasing chitinase and peroxidase activity in rice act as inducing talc-based systemic resistance [33]. А bioformulation reduce the incidence of sheath blight in rice by up to 62%, with 12-21% more grain yield because of trigger the ISR against the microbes [34,35].

# 3.8 PGPR and interference in Quorum sensing

The bacterial cell to cell communication system between themselves or external eukaryotes cues coordinated bacterial chemical the behaviors in response to fluctuation in cell population density called quorum sensing. The small chemical molecules like AHL and HAQs are acts as autoinducer the networking passage communication to enhance the for their concentration as a function of cell density .After their release several bacterial pathogen collectively express the traits like biofilm production ,virulence build up ,swarming motility to protect them against the harsh environmental condition or phagocytosis and hyperparasitism. Some bacterial strains act as interference in biofilm production like Burkholderia glumae, Psudomonas aeruginosa and Bacillus cereus in rice so pathogen virulence diminishes and they acts as the biocontrol agent for the crop [36].

#### 4. CONCLUSION

From above all various information we concluded that in sugarcane and rice PGPR promote plant growth directly and in directly by increasing uptake and availability of macronutrients (N, P, K and essential minerals) and play a role by regulating plant hormone levels indirectly with reducing the inhibitory effects of various pathogens on plant growth and development in the form of biological control agents [33,34,35]. For future need of mankind with hygienic and residue free organic harvest we require careful management without burning the farm residue, mulching, heap management for harboring them in field. They are also act as the biocontrol agent against many disease in both the crop [37]. By active application of microbial consortia at different growth stages of crop eliminate the dependency of big imports of potassic and phosphatic fertilizers with saving the incredible foreign currency . The harmonize and symbiotic management of them must emphasized for satat vikas lakshy in swarnim Bharat with biosphere and ecological conservation having sustainability for our mankind [37,37-41].

### 5. FUTURE SCOPE OF STUDY

In today's climate changing scenario new hybrids varieties are developed only for quantitative higher yield without much bother about the devastating effect on the soil and its loss in productivity . By adopting the application of different PGPR bacterial inoculant in different crop as seed treatment ,seed priming, seedling drenching or spraying the endophytic bacterial culture on standing crop built them healthy and prosperous growth and development. They play pivot role as microbial control, nitrogen fixation, Phosphate solubilization, Potash mobilizing, siderophore formation, different enzymes and auxins formation leads to direct and indirect benefit to plants .By its application we save our foreign currency for importing the chemical fertilizers from international markets. Require better practical approach, wide extension programs, laboratory facility near the village area high shelf and transport life with better CFU count are essential for its wide adaptations. Scope for collecting much information about crop ecological and rhizosphere system with new strains availability of PGPR with their complex interactions in respective ecological and climatic region. They are helpful to farmer to indiscriminate application reduce the of fertilizers, fungicides with enhance the bio safety approach required for today's environment.

#### ACKNOWLEDGEMENT

We thank to our guide DR. Patel, Dr. Rakholiya, Dr. John, DR. Patel from Department of Plant

Pathology, NAU, Navsari (Gujarat) for their valuable suggestion and practical approach with feedback of the topic on earlier drafts of this manuscript.

#### **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

### REFERENCES

- Doni F, Suhaimi NS, Mispan MS, Fathurrahman F, Marzuki BM, Kusmoro J, Uphoff N. Microbial contributions for rice production: From conventional crop management to the use of 'omics' technologies. International Journal of Molecular Sciences. 2022;23(2):737.
- Grifoni A, Bazzicalupo M, Serio CD, Farcelli S, Fani R. Identification of *Azospirillum* strains by restriction fragment length polymorphism of 16 S rDNA and the histidine operon. FEMS Microbiol Lett. 1995;127:85–91.
- Rosa PAL, Galindo FS, Olivera CED, Jalal 3. A, Martinao ES, Fernadas GC, Morgana with plant ER. Inoculation arowthpromoting bacteria to reduce phosphate fertilization requirement and enhance technological quality and yield of sugarcane. Microorganisms. 2022;10(1):19 2 -200.
- Mendes R, Pizzirani-Kleiner AA, Araujo WL, Raaijmakers JM. Diversity of cultivated endophytic bacteria from sugarcane: Genetic and biochemical characterization of *Burkholderia cepacia* complex isolates. Appl Environ Microbiol. 2007;73:7259–7267.
- Arencibia AD, Vinagre F, Estevez Y, Bernal A, Perez J, Cavalcanti J, Santana I, Hemerly AS. *Gluconacetobacter diazotrophicus* elicits a sugarcane defense response against a pathogenicbacteria *Xanthomonas albilineans*. Plant Signal Behav. 2006;1(5):265–273

 Tayade AS, Geetha P, Anusha S, Dhanapal R, Hari K. Bio-intensive modulation of sugarcane ratoon rhizosphere for enhanced soil health and sugarcane productivity under tropical Indian condition. Sugar Tech. 2019;21:278–288.

7. Yanni YG, Rizk RY, El-Fattah FKA, Squartini A, Corich VGA, De Bruijn F, Rademaker J, Maya Flores J, Ostrom P, Vega-Hernandez M, Hollingsworth RI, Martinez-Molina E, Mateos P, Velazquez E, Wopereis J, Triplett E, Umali-Garcia M, Anarna JA, Rolfe BG, Ladha JK, Hill J, Mujoo R, Ng PK, Dazzo FB. The beneficial plant growth-promoting association of *Rhizobium leguminosarum* bv. *trifolii* with rice roots. Aust. J. Pl. Physiol S. 2001;28:845–870.

- 8. Kumar NR, Arasu VT, Gunasekaran P. Genotyping of antifungal compounds producing plant growth-promoting rhizobacteria, *Pseudomonas fluorescens*. Curr Sci. 2002;82(12):1463–1466.
- 9. Viswanathana R, Samiyappan R. Induced systemic resistance by *Pseudomonads flurosence* against red rot disease of sugarcane caused by *Colletotrichum falcatum*. Crop Prot. 2002;21:1–10.
- 10. Antwerpen TV, Rutherford RS, Vogel JL. Assessment of sugarcane endophytic bacteria and *rhizospheric Burkholderia* species as antifungal agents. Proc. S. Afr. Sug. Technol. Assoc. 2002;76:301–304.
- 11. Feng YD, Shen D, W Song. Rice endophyte *Pantoea agglomerans* YS 19 promotes host plant growth and affects allocations of host photosynthates. J. Appl. Microbiol. 2006;100:938-945.
- Chebotar VK, Malfanova NV, Shcherbakov AV, Ahtemova GA, Borisov AY, Lugtenberg B, Tikhonovich IA. Endophytic bacteria in microbial preparations that improve plant development (review). Appl. Biochem. Microbiol. 2015;51:271–277.
- 13. Khin TW, Odd AZ, Ohstu NO, Yokohoma T. *Bacillus pumilis* strain TUAT - and nitrogen application in nursery phase promote growth of rice plants under fields condition. Agronomy. 2018;8(10):216-222.
- 14. Abdelaziz S, Hemeda NF, Belal EE, Elanberg R. Efficacy of facultative oligotrophic bacterial strain as PGPR and their potency against two pathogenic fungi causing damping off disease. Applied microbial. Open access; 2018.
- 15. Seenivasan N. Efficacy of *Pseudomonas fluorescens* and *Paecilomyces lilacinus* against *Meloidogyne graminicola*
- 16. infesting rice under system of rice intensification. Arch. Phytopathol. Plant. Prot. 2011;44:1467–1482.
- Biswas JC, Ladha JK, Dazzo FB, Yanni YG, Rolfe BG. Rhizobial inoculation influences seedling vigor and yield of rice. J. Agron. 2000;92:880–886.
- 18. Isawa T, Yasuda M, Awazaki H, Minamisawa K, Shinozaki S, Nakashita H.

*Azospirillum* sp. strain B510 enhances rice growth and yield. Microbes Environ. 2010; 25:58–61.

- Ghosh A, Pramanik K, Bhattacharya S, Mondal S, Ghosh SK, Ghosh PK, Maiti TK. Abatement of arsenic-induced phytotoxic effects in rice seedlings by an arsenicresistant *Pantoea dispersa* strain. *Environ*. Sci. Pollut. Res. 2021;28:21633–21649.
- 20. Patel A, Kumar A, Sheoran N, Kumar M. Sahu KP. Antifungal and defense elicitor activities of pyrazines identified in endophytic *Pseudomonas putida* BP25 against fungal blast incited by *Magnaporthe oryzae* in rice. J. Plant Dis. Prot., 2021;128:261–272.
- 21. Oliveira MIDS, Chaibub AA, Sousa TP, Cortes MV, D. Souza AC, Conceicao ECD, Filippi MC. Formulations of *Pseudomonas fluorescens* and *Burkholderia pyrrocinia*
- 22. control rice blast of upland rice cultivated under no-tillage system. Biol. Control. 2019 ;27:104-153.
- 23. Chan C, Zhu H. Are common symbiosis gene required for endophytic rice rhizobial interaction. Plant Signaling Behavior. 2013;8(9):25453.
- Alam MS, Cui ZJ, Yamagishi T, Ishii R. Grain yield and related physiological characteristics of rice plants (*Oryza* sativa L.) inoculated with free-living rhizobacteria. Plant Prod. Sci. 2001;4:126– 130.
- 25. Yanni YG, Rizk RY, Corich V, Squartini A, Ninke K, Philip-Hollingsworth S. Natural endophytic association between *Rhizobium leguminosarum* bv. *trifolii* and rice roots and assessment of its potential to promote rice growth. Plant Soil.1997;194:99–114.
- 26. Biswas JC, Ladha JK, Dazzo FB. Rhizobia inoculation improves nutrient uptake and growth of lowland rice. Soil Sci. Soc. Am. J. 2009;64:1644–1650.
- Leelasuphakul W,Sivanunsakul P,and Phogpaichit S, Purification ,characterization and synergetic activity of *beta* 1 3 gulcanase and antibiotics extract from an antagonistic Bacillus subtillis NSRS 89-24 against rice blast and sheath blight .Enzyme Micro Technol.38:990-997.
- 28. Durgadevi D, Harish S, Manikandan R, Prabhukarthikeyan SR, Alice D. Raguchander T. Proteomic profiling of defense/resistant genes induced during the tripartite interaction of Oryza sativa, Rhizoctonia solani AG1-1A, and Bacillus subtilis against rice sheath

blight. Physiol. Mol. Plant Pathol. 2021; 11(5):101-669.

- Yang D, Wang B, Wang J, Chen Y, Zhou M. Activity and efficacy of *Bacillus* subtilis strain NJ-18 against rice sheath blight and *Sclerotinia* stem rot of rape. Biol. Control. 2009;51:61–65.
- Someya N, Nakajima M, Watanabe K, Hibi T, Akutsu K. Potential of Serratia marcescens strain B2 for biological control of rice sheath blight. Biocontrol Sci. Technol. 2005;15:105–109.
- 31. Stockwell VO, Stack JP. Using *Pseudomonas* spp. for integrated biological control. Phytopathology. 2007;9 7:244–249.
- 32. Nandakumar R, Babu S, Viswanathan R, Raguchander T, Samiyappan R..Induction of systemic resistance in rice against sheath blight disease by *Pseudomonas fluorescens*. Soil Biol. Biochem. 2001;33:6 03–612.
- Radjacommare R, Nandakumar R, Kandan A, Suresh S, Bharathi M, Raguchander T, Samiyappan R. *Pseudomonas fluorescens* based bioformulation for the management of sheath blight and leaf folder in rice. Crop Prot. 2002;21:671–677.
- 34. Vidhyasekaran Ρ. Rabindran R, Nayar Muthamilan Μ, Rajappan Κ, Subramanian Κ, Vasumathi K. Development of powder formulation of Pseudomonas fluorescens for control of rice blast. Plant Pathol. 1997;46:291-297.
- 35. Paluch E. Soroczynskra JR, Jedrisik I, Mazurkiewicz E, Jermakow K. Prevention

of biofilm formation by quorum quenching. Applied Microbiology and Biotechnology. 2020;104:1871-1881.

- Tariq M, Noman M, Ahmed T, Hameed A, Manzoor N, Zafar M. Antagonistic features displayed by plant growth promoting rhizobacteria (PGPR): A review. J. Plant Sci. Phytopathol. 2017;1:38–43.
- Blanco Y, Pinon D, Legaz ME, Vicente C. Antagonism of *Gluconacetobacter diazotrophicus* (a sugarcane endosymbiont) against *Xanthomonas albilineans* (pathogen) studied in alginate-immobilized sugarcane stalk tissues. J. Biosci. Bioeng. 2005;99:366-371.
- Alves BJR, Reis V. Endophytic nitrogen fixation in sugarcane: Present knowledge and future applications. Plant Soil. 2003;252:139-149.
- 39. Hassan MN, Afghan S, Hafeez FY. Biological control of red rot in sugarcane by native pyobutine producing *P. putida* strain NH 50 under field condition and its potential mode of action Pest Management Sci. 2011;67(9):1147-1154..
- 40. Khandagale PP, Kesker MM, Ghodke SK, Rasker BS. Study the effect of the *Gluconoacetobacter diazotrophicus* and PSB formulation on quality parameter on suru sugarcane. Int. J. Current Microbiology Applied Sci. 2019;8(5):210-2 15.
- 41. Pedraza RO. Recent advances in nitrogenfixing acetic acid bacteria. Int. J. Food Microbiol. 2008;125(1):25–35.

© 2024 Khandagale et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle5.com/review-history/112803