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# **Enhancement of Genetic Variability for Yield and Component Traits through Recombination followed by Induced Mutagenesis in Greengram [***Vigna radiata* **(L.) Wilczek]**

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## *Authors' contributions*

*This work was carried out in collaboration among all authors.The experiment was performed and the manuscript was written by author PKM. Author SCM supervised the experiment and revised the manuscript. Author LSA helped in taking observations and data analysis. All authors read and approved the final manuscript.*

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*Original Research Article*

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# **ABSTRACT**

**Aim:** Greengram is a self-pollinated crop which show very less variability to develop improved varieties through only hybridization or induced mutation breeding. Therefore, we have taken a new pace to create more variability by combining both recombinations with induced mutation through gamma rays irradiation. For this purpose, the  $F_2$  seeds were irradiated with gamma rays at BARC, Mumbai and sown to grow the  $F_2M_1$  generation and subsequently the superior mutant lines with high degree variability with high GCV and genetic advances were selected from  $F_2M_2$  generation of the mutant population.

**Methodology:** The present investigation was carried out during *kharif*-2017 and *rabi-summer* 2017-18 at the experimental plot, All India Coordinated Research Projects (AICRP) on MULLaRP,

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main Agricultural Research Station, College of Agriculture, University of Agricultural Sciences, Dharwad, Karnataka, India. The experiment was laid out in an augmented design.

**Results:** The mutant progenies obtained from the crosses DGGV-2 × IPM-410-3 and DGGV-2 × SML-1815 in  $F_2M_2$  generation have shown high PCV and GCV for the characters like plant height, number of clusters per plant and number of seeds per pod etc. when irradiated with 100 kR gamma rays. The mutant breeding lines derived from the crosses DGGV-7 × V-02-709 and DGGV-7 × V-02-802 with irradiation dose of 20 kR, have shown higher number of pods per cluster and higher number of pods per plant with high heritability. More variability was observed with higher dose (100 kR) of mutation even though it showed higher mortality rate.

**Conclusion:** Irradiation of F<sub>2</sub> progeny (DGGV-2 × SML-1815) with 100 kR has generated more genetic variability for seed yield per plant (10.8 g), when compared to the check DGGV-2 (4.7 g) and SML-1815 (9.8 g). So, priority should be given to those characters which are having high heritability coupled with high genetic advance as per cent mean to get better selection gains. The breeding lines which showed higher degree of variability can be utilized in the future breeding programme for development of high yielding genotypes.

*Keywords: Variability; heritability; genetic advance; mutation; gamma rays.*

## **1. INTRODUCTION**

Greengram [*Vigna radiata* (L.) Wilczek, 2n=2x=22] is an important grain legume among the major pulses with approximately 25-28% protein, 62-65% carbohydrates and 3.5-4.5% fibre on dry weight basis [1]. This crop is cultivated in the tropical and sub-tropical regions of the country mostly as a part of cereal based cropping system [2]. This crop is popular among farmers due to its early maturity, drought tolerance and ability to fix biological Nitrogen  $(30-40 \text{ kg} \text{ ha}^{-1})$  which allows it to thrive in Nitrogen deficit soils [3]. Yield is a complex trait which is influenced by several factors including the environmental influence. For this reason critical selection of the yield attributing traits in greengram is important. The selection process mainly depends on the extent and amount of genetic variability present in the available breeding materials [4]. The genetic variability in greengram, is drastically reduced during the course of evolution as it is autogamous in nature [5]. There are many attempts have been made to develop improved varieties through hybridization and selection, but the yield potentiality of greengram remains static due to shortage of enough variability [6,7].

Mutation breeding is relatively easier method of crop improvement which is mainly based on the conventional breeding approach which brings novel genotypes with high yielding ability through heritable genetic changes of specific traits. A number of research evidences are available on the genetic variability study on greengram segregating materials by [8,9,10] and mutagenic variability study on different adopted varieties to

create variation by several researchers [11,12] on greengram. But, it is realized that the variability developed through hybridization or by induced mutation alone is not sufficient to select the desirable variants from the population due to self-pollination nature. Thus, in the present investigation we had taken an innovative step to create sufficient amount of desirable variability in greengram by ameliorating both hybridization and induced mutation through gamma rays. Therefore, the segregating materials  $(F<sub>2</sub>$  seeds) were treated with gamma rays to create high variability through recombination followed by induced mutagenesis. There is a chance for improving variability in agro-morphological traits for selecting superior lines to increase their productivity and quality in greengram. With this aspiration the present investigation aimed at induction of mutation in segregating lines of greengram using gamma irradiation. The genotypic and phenotypic variability (GCV and PCV), heritability and genetic advance (GA) for yield components were assessed and the superior single cross progeny lines were selected in  $F_2M_2$  generation of the mutant population.

#### **2. MATERIALS AND METHODS**

The field experiment was carried out during *kharif*-2017 and *rabi-summer* 2017-18 in an augmented design at Experimental plot, AICRP on MULLaRP, Main Agricultural Research Station, College of Agriculture, University of Agricultural Sciences, Dharwad, Karnataka, India. The  $F<sub>2</sub>$  (Healthy and well dried) seeds derived from the following four different crosses mentioned in Table 1; were sent to Bhabha Atomic Research Centre, Trombay, Mumbai for gamma rays irradiation in *kharif*-2017 for the creation of desirable variability. Even though the  $LD_{50}$  for germination and survival of the seedlings range between 40-50 kR of gamma rays in greengram [7] but, 20-60 kR also reported [13,14]. In this study we have treated with 20 and 100 kR of gamma rays. Even though use of 100 kR is not usually practiced, but we have treated our materials expecting to get some novel mutants from the breeding lines. The pedigree, number of seeds from each cross and the dose of irradiation are stated below in Table 1.

# **2.1 Experimental Methodology**

The gamma rays irradiated seeds were sown in augmented design during *kharif-*2017 along with their respective checks to grow the  $F_2M_1$ generation. The individual plants were critically observed from the date of germination to the maturity. The desired variability on plant growth (7), robustness (4), maximum number of flowering branches (5), pod length (10) and seed size (5) etc. were observed in the respective number of plants and they were selected as putative mutants (as expression of dominant mutants). The mutant lines are compared with the parental lines and also with the  $F_3$  progeny lines derived from the following crosses as check. The putative mutants selected and tagged in  $F_2M_1$  and the seeds were harvested separately to develop the  $F_2M_2$  mutant population in *summer*-2018. The population size maintained in  $F<sub>2</sub>M<sub>2</sub>$  generation was nearly 1000 plants for each of the cross derivatives. The morphological observations for the traits mentioned in Table 2; were taken in  $F_2M_2$  generation to study the genetic variability.

The phenotypic and genotypic coefficient of variability (PCV and GCV) for all the characters was estimated using the formulae of Burton and De Vane [15]. The GCV and PCV values were categorized as low (0-10%), moderate (10-20%) and high (> 20%) as indicated by Shiva Subramanian and Menon [16]. Heritability (broad sense) was estimated for all the characters as the ratio of genotypic variance to the total variance as suggested by Lush [17] and Hanson [18]. According to Robinson [19] heritability estimates in cultivated plants can be placed in the categories; low (0-30%), moderate (30-60%) and high (> 60%). Genetic advance for each character was estimated by using the formula of Johnson [20] and Genetic advance as per cent of mean (GAM) was categorized according to him as low (0-10%), moderate (10-20%) and high  $(20\%).$ 

# **3. RESULTS AND DISCUSSION**

The analysis of variances revealed significant variability for most of the characters in all the four mutagen treated breeding lines (Table 2). The mutant progenies of the cross derivative of DGGV-7 × V-02-709 has shown significant variation for the characters like days to 50 per cent flowering, number of pods per cluster, 100 seed weight (g) and seed yield per plant (g) with dose of 20 kR. However, significant variation was seen with 100 kR for days to plant height, number of clusters per plant, pod length, 100 seed weight and seed yield per plant for derivatives of DGGV-2 × SML-1815; whereas, non-significant variation was seen for number of seeds per pod with 100 kR. The magnitude of shift in the mean value varied with the dose of mutagen and the parent materials. Similar findings also reported by [21] in greengram.

# **3.1 Estimates of Variability (GCV and PCV), Heritability and Genetic Advance (GA) for Yield Components in F2M2 Generation**

Depending upon the magnitude of genetic variability in different treatment populations, the genetic parameters like GCV, PCV, heritability and genetic advance under selection was varied. These helps in induction of micro and macro mutations and thereby increasing the scope of improvement for the desirable traits through selection. The genetic variability parameters for different yield attributing characters are presented in the Table 3.







# **Table 2.ANOVA for yield and component traits of green gram breeding lines in F2M2 generation**

*Where, C1- DGGV-7 × V-02-709 (20 kR), C2- DGGV-7 × V-02-802 (20 kR), C3- DGGV-2 × IPM-410-3 (100 kR), C4- DGGV-2 × SML-1815 (100 kR). The values within parenthesis along with the crosses indicate the doses of Gamma rays irradiation. \* and \*\* - Significant at 5 % and 1 % level of probability respectively*

# **3.1.1 Days to 50% flowering**

Days to 50 per cent flowering was observed in advance from the mutant breeding lines derived

from the cross DGGV-7 × V-02-709 when treated with 20 kR gamma rays as compared to nonmutagenized lines. The average days to 50 per cent flowering was showed 41.32 days with the range between minimum and maximum days to 50 per cent flowering of 36.63 and 44.36 days. Highest heritability was also observed for the progenies of the same cross, coupled with higher genetic advance over mean. Higher PCV (3.98%) and GCV (2.42%) was observed for the cross derivative of DGGV-2 × IPM-410-3 with irradiation dose of 100 kR as compared to the other derivatives with 20 kR.

## **3.1.2 Number of pods per plant**

The heritability 65.23% was observed for the cross derivative of DGGV-7 × V-02-802 when treated with 20 kR and it has also recorded higher mean number of pods per plant. The phenotypic coefficient of variability was recorded 57.24% in the cross derivative of DGGV-2 × SML-1815, but the heritability was moderate when treated with 100 kR.

#### **3.1.3 Number of seeds per pod**

Higher number of seeds per pod *i.e*. 12.22 was recorded in the cross derivative of DGGV-7 × V-02-802 when treated with 20 kR. The higher heritability (12.30%) with genetic advance over mean was recorded in these breeding mutant lines. Moderate GAM (10.32%) with high heritability (70.23%) was observed for the progeny of DGGV-2 × IPM-410-3.

## **3.1.4 100 seed weight**

The mutant progenies derived from the cross DGGV-2 × SML-1815, after treatment with 100 kR g showed high PCV (23.45%) but GCV (12.64%) was low for the same cross. Moderate genetic advance under mean (18.46%) with 74.36% of heritability was shown in the mutant lines derived from the cross DGGV-7 × V-02-802 with 20 kR.

## **3.1.5 Seed yield per plant**

The maximum range of seed yield per plant (6.95 to 23.65 g) was observed in the cross derivatives of DGGV-2 × IPM-410-3 (100 kR) with PCV (38.69%) and GCV (21.36%). The progeny lines derived from the cross DGGV-2 × SML-1815 (100 kR) has shown moderate heritability (59.87%) and high genetic advance over mean (48.96%).

The variability parameters are compared between the normal segregating population in  $F_3$ generation [22] and in  $F_2M_2$  generation on the same cross derivatives. It was observed higher variability for most of the yield attributing traits in greengram in  $F_2M_2$  generation as compared to  $F_3$ generation. In the present investigation, there was a reduction of yield per plant in higher doses, but the seed size increased with the increased dose of mutagen as compared to the checks. The increase in the variance of  $F_2M_2$ population in a trait is a general indicator of induction of micro mutation with negative and/or positive on the trait. Similar trend in increasing variability in  $M_2$  populations with different doses of mutagens on various yield attributing characters in greengram was earlier reported by [6,21,23]. The number of clusters per plant was increased with dwarf plant type was noticed by increased dose of mutagen [24]. Whereas, some reverse results also reported with increased dose of mutagen for seed yield per plant [25] and plant height in greengram [26]. The results reported by many other researchers on GCV and PCV are comparable with the present investigations on effect of gamma rays on greengram [27,28,29]. The heritability estimates along with genetic advance is usually more helpful than the heritability value alone in selected lines [20]. The genetic advance is an indicative of the expected genetic progress for a particular trait under suitable selection procedure. The earlier findings on heritability for different traits in greengram were also similar with the present study [13,14,29,30].

# **3.2 Evaluation of Irradiated Single Cross**  Progenies (F<sub>2</sub>M<sub>2</sub>) for Yield and **Component Traits during** *Summer***-2018**

The progeny line numbers 1M-2 and 1M-5 derived from DGGV-7 × V-02-709 treated with 20 kR gave a seed yield of 7.2 g per plant. Number of pods per plant produced by these lines were 27 and 26 respectively. Seed yield per plant of 5.3 and 5.7 g were recorded for the progeny numbers 1M-14 and 1M-17. The progeny lines derived from the cross DGGV-7 × V-02-709 irradiated with 20 kR dose, revealed earliness indicated by the characters days to flowering (37 days) and maturity (84 days). 100 seeds weight of 2.6 g was recorded for the progeny numbers 2M-4, 2M-6 and 2M-18. Higher seed yield per plant was recorded 7.4 and 7.2 g in the progeny lines of 2M-11 and 2M-18 respectively from the cross derivatives of DGGV-7 × V-02-802 with 20 kR. It was recorded 6.7 and 8.3 g of seed yield per plant for the progeny number 3M-9 and 3M-15 respectively derived from the cross DGGV-2 × IPM-410-3 when treated with 100 kR gamma rays.

SI.	<b>Characters</b>	<b>Crosses</b>	<b>Parental</b>	Progeny	Range		<b>PCV</b>	<b>GCV</b>	$h^2$ <sub>bs</sub>	GA	<b>GAM</b>
no.			mean	mean	<b>Maximum</b>	<b>Minimum</b>	(%)	(%)	(%)		$(\%)$
$\mathbf{1}$ .	Days to 50 %	C <sub>1</sub>	41.24	41.32	36.63	44.36	3.65	2.29	28.33	0.57	1.39
	flowering	C <sub>2</sub>	41.32	42.13	37.99	43.38	2.25	1.44	27.25	0.50	1.21
		C <sub>3</sub>	41.25	45.33	39.25	48.23	3.98	2.42	25.36	0.44	1.09
		C <sub>4</sub>	40.00	44.29	39.45	47.17	3.74	2.16	24.55	0.41	1.03
$\overline{2}$ .	Plant height	C <sub>1</sub>	58.22	51.24	40.12	69.44	29.46	23.54	69.05	18.67	36.44
	(cm)	C <sub>2</sub>	57.50	54.28	40.22	71.46	32.22	27.13	65.22	18.59	34.26
		C <sub>3</sub>	52.34	56.32	42.35	72.65	36.41	28.31	54.63	13.69	24.31
		C <sub>4</sub>	52.5	55.26	44.63	72.55	35.54	26.25	55.36	15.67	28.36
$\overline{3}$ .	Number of	C <sub>1</sub>	4.5	5.28	3.82	6.19	$\overline{26.95}$	21.66	46.23	1.76	33.45
	branches per plant	C <sub>2</sub>	4.0	5.15	3.77	6.75	28.15	22.25	55.12	1.87	36.36
		C <sub>3</sub>	4.26	5.69	3.88	7.42	32.65	31.22	78.24	3.09	54.32
		C <sub>4</sub>	5.75	6.21	3.79	7.16	33.74	30.16	76.63	3.18	51.22
4.	Number of clusters	C <sub>1</sub>	8.00	7.44	5.16	17.12	39.46	24.46	88.36	4.85	65.32
	per plant	C <sub>2</sub>	7.5	7.76	5.46	18.12	39.29	23.11	82.22	4.83	62.31
		C <sub>3</sub>	8.65	8.66	5.49	20.31	43.63	17.35	68.21	4.44	51.36
		C <sub>4</sub>	8.54	8.83	4.99	18.46	42.69	16.22	64.23	4.32	48.95
5.	Days to 50 %	$\overline{C1}$	41.24	41.32	36.63	44.36	3.65	2.29	28.33	0.57	1.39
	flowering	C <sub>2</sub>	41.32	42.13	37.99	43.38	2.25	1.44	27.25	0.50	1.21
		C <sub>3</sub>	41.25	45.33	39.25	48.23	3.98	2.42	25.36	0.44	1.09
		C <sub>4</sub>	40.00	44.29	39.45	47.17	3.74	2.16	24.55	0.41	1.03
6.	Plant height	$\overline{C1}$	58.22	51.24	40.12	69.44	29.46	23.54	69.05	18.67	36.44
	(cm)	C <sub>2</sub>	57.50	54.28	40.22	71.46	32.22	27.13	65.22	18.59	34.26
		C <sub>3</sub>	52.34	56.32	42.35	72.65	36.41	28.31	54.63	13.69	24.31
		C <sub>4</sub>	52.5	55.26	44.63	72.55	35.54	26.25	55.36	15.67	28.36
$\overline{7}$ .	Number of	$\overline{C1}$	4.5	5.28	3.82	6.19	26.95	21.66	46.23	1.76	33.45
	branches per plant	C <sub>2</sub>	4.0	5.15	3.77	6.75	28.15	22.25	55.12	1.87	36.36
		C <sub>3</sub>	4.26	5.69	3.88	7.42	32.65	31.22	78.24	3.09	54.32
		C <sub>4</sub>	5.75	6.21	3.79	7.16	33.74	30.16	76.63	3.18	51.22
8.	Number of clusters	$\overline{C1}$	8.00	7.44	5.16	17.12	39.46	24.46	88.36	4.85	65.32
	per plant	C <sub>2</sub>	7.5	7.76	5.46	18.12	39.29	23.11	82.22	4.83	62.31
		C <sub>3</sub>	8.65	8.66	5.49	20.31	43.63	17.35	68.21	4.44	51.36
		C <sub>4</sub>	8.54	8.83	4.99	18.46	42.69	16.22	64.23	4.32	48.95

Table 3.Estimate of variability, heritability and genetic advance for yield components in F<sub>2</sub>M<sub>2</sub>

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Where, PCV- Phenotypic coefficient of variation, GCV- Genotypic coefficient of variation, h<sup>2</sup>(bs)- Broad sense heritability, GA- Genetic advance, GAM- Genetic advance as per *cent mean, C1- DGGV-7 × V-02-709 (20 kR), C2- DGGV-7 × V-02-802 (20 kR),C3- DGGV-2 × IPM-410-3 (100 kR), C4- DGGV-2 × SML-1815 (100 kR). The values within parenthesis along with the crosses indicate the doses of Gamma rays irradiation*



Table 4. Evaluation and selection of irradiated single cross progenies (F<sub>2</sub>M<sub>2</sub>) for yield and yield components during summer-2018

26, 24 and 36 number of pods per plant were observed for the progeny numbers 3M-9, 3M-10 and 3M-15 respectively (Table 4).

Higher seed yield per plant and 100 seed weight was recorded in the progeny number 4M-9 and 4M-18 derived from DGGV-2 × SML-1815 when treated with 100 kR gamma rays. The seed yield per plant was 8.3 g and 10.8 g respectively; while the 100 seed weight was recorded as 5.4 g for both the progenies. These findings were in accordance with the earlier reports for seed yield [21,22,31,32]. From the present study, days to 50 per cent flowering and days to maturity was delayed with 100 kR dose as compared to 20 kR. But, 100 seed yield per plant was increased with 100 kR with respect to 20 kR. There are some rare alleles present in the conserved gene block, but they are usually not expressed. The result observed here may be due to these novel alleles which are expressed through higher dose of mutation. The present findings showed conformity with the results reported earlier only with irradiation [33,34,35]. If the parents have higher variability then there is a chance that the progenies will also express more variability and it will be helpful for direct selection of the traits [36]. In the present investigation, recombination followed by irradiation with gamma rays gave a better result as compared to only recombination or only mutation. The traits which revealed sufficient variability with high amount of heritability and genetic advance will favor for selection of superior recombinants in further generation.

# **4. CONCLUSIONS**

Induced mutagenesis followed by recombination offers an opportunity to express the hidden variability in the conserved gene blocks which is very pertinent to green gram for creation of desirable variability as it is self-pollinated crop. Although, 40 kR is a lethal dose of gamma irradiation, in the current investigation irradiation of  $F<sub>2</sub>$  progeny (DGGV-2  $\times$  SML-1815) with 100 kR has generated more genetic variability for seed yield per plant (10.8 g), when compared to the check DGGV-2 (4.7 g) and SML-1815 (9.8 g). Further four superior progenies *viz*; 4M-5, 4M-9, 4M-18 and 4M-20 have recorded highest 100 seed weight 7.9, 8.3, 10.8 and 7.9 g respectively. Thus, it can be inferred that high yielding  $F_2M_2$ progenies in the mutagenized populations with significant mean yield would be effective for the selection of desirable high yielding micro/macro mutants. The higher dose of gamma rays in

greengram will provide enough scope to develop a wide range of variation in desirable plant attributes which may facilitate to select high yielding mutants with other desirable characteristics like long pod with larger seed size, early maturity, short-statured plant etc. These superior genotypes can be used in further breeding programmes by introgressing all the desirable traits. Further analysis of the morphological macro/micro mutants can be differentiated from the normal plants through marker assisted selection.

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# **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

## **REFERENCES**

- 1. Gowda CL, Laxmipathi, Sushil K, Chaturvedi, Pooran M, Gaur CV, Sameer K, Aravind KJ. Pulses research and development strategies for India GRSV Consulting Services, ICRISAT, IIPR, CAZRI, India; 2015.
- 2. Nair RM, Yang RY, Easdown WJ, Thavarajah D, Thavarajah P, Hughes J, et al. Biofortification of greengram (*Vigna radiata*) as a whole food to enhance human health. Journal of Science, Food and Agriculture. 2013;93:1805-1813.
- 3. Tak S, Sharma SK, Reager ML. Growth attributes and nutrient uptake of green gram as influenced by vermicompost and zinc in arid Western Rajasthan. Advance Research Journal of Crop Improvement. 2013;2(1):65-69.
- 4. Susmitha D, Jayamani P. Genetic variability studies for yield and its contributing traits in greengram [*Vigna radiata* (L.) Wilczek]. Electronic Journal of Plant Breeding. 2018;9(2):716-722.
- 5. Singh SK, Lavanya GR, Bhat KV, Babu GS, Arya L, Verma M, Hussain Z, Roy S, Rathi RS, Mishra AK. Microsatellite

markers revealed genetic diversity in greengram mutant lines. Indian Journal of Hill Farming. 2012;25(1):38-43.

- 6. Kozgar MI, Goyal S, Khan S. EMS induced mutational variability in *Vigna radiata* and *Vigna mungo*. Research Journal of Botany. 2011;6:31-37.
- 7. Rukesh AG, Abdul Rahuman M, Latitia SC, Packiaraj D. Impact of gamma irradiation induced mutation on morphological and yield contributing traits of two genotypes of Green gram (*Vigna radiata* L.). Journal of Pharmacognosy and Phytochemistry. 2017;6(6):1229-1234.
- 8. Aalok S, Vinita R, Vadodariya GD, Modha KG, Patel RK. Genetic variability, heritability and genetic advance in  $F_3$ progenies of greengram [*Vigna radiata* (L.) Wilczek]. International Journal of Current Microbiology and Applied Sciences. 2017;6(12):3086-3094.
- 9. Pulagampalli R, Roopa GL. Variability, heritability genetic advance and correlation coefficients for yield component characters and seed yield in Greengram (*Vigna radiate* (L.) Wilczek). Journal of Pharmacognosy and 2017;6(4):1202-1205.
- 10. Reddy DKR, Venkateswarlu O, Obaiah MC, Jyothi GLS. Studies on genetic variability, character association and path coefficient analysis in greengram. Legume Research. 2011;34(3):202-206.
- 11. Auti SG. Lhb mutant- A novel mutant of greengram [*Vigna radiata* (L.) Wilczek] induced by gamma radiation. Bioremediation, Biodiversity and Bioavailability. 2012;6(1):87-93.
- 12. Usharani KS, Anandakumar CR. Estimation of variability, heritability and genetic advance in mutant populations of black gram [*Vigna mungo* (L.) Hepper]. SABRAO Journal of Breeding and Genetics. 2016;48(3):258-265.
- 13. Das TR, Baisakh B. Mutation-induced polygenic variability and early prediction of high yielding mutants in greengram [*Vigna radiata* (L.) Wilczek]. International Journal of Current Microbiology and Applied Sciences. 2018;7(1):3228-3236.
- 14. Mishra D, Singh B, Sahu R. Gamma ray induced macro-mutations in green gram [*Vigna radiata* (L.) Wilczek]. Internatioal Journal of Agriculture and Forestry. 2013;3(3):105-109.
- 15. Burton GW, De Vane EM. Estimating heritability in tall fescue (*Festuca*

*arundinaceae*) from replicated clonal material. Agronomy Journal. 1953;51(5): 515-518.

- 16. Sivasubramanian S, Menon M. Heterosis and inbreeding depression in rice. Madras Agriculture Journal. 1973;60(5):1339.
- 17. Lush JL. Heritability of quantitative characters in farm animals. Proc. 85<sup>th</sup> Cong. Genetics. 1949;356-375.
- 18. Hanson GH, Robinson HF, Comstock RE. Biometrical studies of yield in segregating populations of Koren Lependeza. Agronomy Journal*.* 1956;48(1):267-282.
- 19. Robinson HF, Comstock RE, Harvey PH. Estimation of heritability and degree of dominance in corn. Agronomy Journal. 1949;41(3):353-359.
- 20. Jhonson HW, Robinson HI, Comstock RE. Estimation of genetic and environmental variability in soybean. Agronomy Journal. 1955;47(3):314-318.
- 21. Majhi PK, Mogali SC. Studies on mutagenic effectiveness and efficiency of gamma rays in greengram [*Vigna radiata* (L.) Wilczek]. International Journal of Current Microbiology and Applied Sciences. 2020;9(3):1475-1484.
- 22. Majhi PK, Mogali SC, Abhisheka LS. Genetic variability, heritability, genetic advance and correlation studies for seed yield and yield components in early segregating lines (F3) of greengram [*Vigna radiata* (L.) Wilczek]. International Journal of Chemical Studies. 2020;8(4):1283-1288.
- 23. Kumar A, Parmhansh P, Kumar R, Prasad<br>R. Induced genetic variability for R. Induced genetic variability for quantitative characters in greengram [*Vigna radiate* (L.) Wilczek]. Indian Journal of Agriculture. 2008;52:93-98.
- 24. Tickoo J. Spectrum and frequency of induced macromutations in greengram. Crop Improvement Society, India. 1987;116-117.
- 25. Ahmad HM, Ahsan M, Ali Q, Javed I. Genetic variability, heritability and correlation studies of various quantitative traits of greengram (*Vigna radiata* L.) at different radiation levels. International Research Journal of Microbiology. 2012;3(11):352-362.
- 26. Khan S, Siddiqui B, Mohammad N, Nadeem M. Variation in quantitative character of greengram after seed treatment with Diethyl sulphate (DES). Advances in Plant Science. 1995;7:41-45.
- 27. Mullainathan L, Gandhi E, Anthoniraju A. Physical and chemical mutagens induced

the mutation in  $M_3$  generation of greengram (*Vigna radiata* L. Wilczek). International Journal of Current Sciences. 2013;6:58-62.

- 28. Mullainathan L, Umavathi S. Induced<br>mutagenesis in *Cicer arietinum*. mutagenesis in *Cicer arietinum*. International Letter of Natural Sciences. 2014;7:1-4.
- 29. Sharma V, Kumar G, Kumar R. EMS induced polygenic variations in greengram. Journal of Food Legumes. 2008;21:61- 62.
- 30. Nair R, Mehta AK. Induced mutagenesis in cowpea [*Vigna ungiculata* (L.) Walp] var. ArkaGarima. Indian Journal of Agricultural Research. 2014;48(4):247-257.
- 31. Das TR, Mishra RC. Genetic analysis of mutagen induced variability in yield traits in greengram (*Vigna radiata*). Environment and Ecology. 2005;23:381-384.
- 32. Desai VK, Parmar LD, Chaudhary AR, Chaudhary NB. Genetic variability, correlation, path coefficient and stability

analysis for yield and its attributing traits in summer greengram [*Vigna radiata* (L.) Wilczek] accessions. International Journal of Current Microbiology and Applied Sciences. 2020;9(6):2942-2955.

- 33. Khan S, Wani MR, Praveen K. Quantitative variability in greengram induced by chemical mutagen. Legume Research. 2006;29(2):143-145.
- 34. Mishra LD, Baishak B, Nayak PK. Genetic variability among advanced mutant lines of greengram. Journal of Food Legumes. 2008;21:274-275.
- 35. Vairam N, Ibrahim SM, Vanniarajan C. Frequency and spectrum of chlorophyll mutations in greengram [*Vigna radiata* (L.) Wilczek]. Asian Journal of Biological Sciences. 2014;9(2):204-207.
- 36. Zuge SS, Abhinav S, Neeraj AT. Genetic variability of yield and yield related traits in greengram [*Vigna radiata* (L.) Wilczek] genotypes. Agric Research Journal. 2019;56(1):163-165.

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